

# The Present and the Future of Degradable Dendrimers and Derivatives in Theranostics

**Victoria Leiro<sup>\*†‡</sup>, João Pedro Garcia<sup>†‡§</sup>, Helena Tomás<sup>⊥</sup>, Ana Paula Pêgo<sup>\*†‡§||</sup>**

<sup>†</sup>INEB–Instituto de Engenharia Biomédica, Universidade do Porto, Rua do Campo Alegre, 823, 4150-180 Porto, Portugal

<sup>‡</sup>Instituto de Investigação e Inovação em Saúde (i3S),

<sup>§</sup>Faculdade de Engenharia da Universidade do Porto (FEUP), and

<sup>||</sup>Instituto de Ciências Biomédicas Abel Salazar (ICBAS) - Universidade do Porto, Porto, Portugal

<sup>⊥</sup>CQM–Centro de Química da Madeira, MMRG, Universidade da Madeira, Campus Universitário da Penteada, 9000-390 Funchal, Portugal

Corresponding Authors: \*E-mail: victoria.leiro@ineb.up.pt.

\*E-mail: apego@ineb.up.pt.

Originally published in: Bioconjugate Chem. 2015, 26, 1182–1197. DOI:10.1021/bc5006224

"This document is the Accepted Manuscript version of a Published Work that appeared in final form in Bioconjugate Chem, copyright © American Chemical Society and the Division of Chemical Education, Inc., after peer review and technical editing by the publisher. To access the final edited and published work see <https://pubs.acs.org/doi/10.1021/bc5006224>."

## ABSTRACT

Interest in dendrimer-based nanomedicines has been growing recently, as it is possible to precisely manipulate the molecular weight, chemical composition, and surface functionality of dendrimers, tuning their properties according to the desired biomedical application. However, one important concern about dendrimer-based therapeutics remains—the nondegradability under physiological conditions of the most commonly used dendrimers. Therefore, biodegradable dendrimers represent an attractive class of nanomaterials, since they present advantages over conventional nondegradable dendrimers regarding the release of the loaded molecules and the prevention of bioaccumulation of synthetic materials and subsequent cytotoxicity. Here, we present an overview of the state-of-the-art of the design of biodegradable dendritic structures, with particular focus on the hurdles regarding the use of these as vectors of drugs and nucleic acids, as well as macromolecular contrast agents.

INSTITUTO  
DE INVESTIGAÇÃO  
E INOVAÇÃO  
EM SAÚDE  
UNIVERSIDADE  
DO PORTO

Rua Alfredo Allen, 208  
4200-135 Porto  
Portugal  
+351 220 408 800  
info@i3s.up.pt  
[www.i3s.up.pt](http://www.i3s.up.pt)

## 1 INTRODUCTION

Natural and synthetic macromolecular structures have frequently been used as vectors for the delivery of drugs and therapeutic nucleic acids, as well as diagnostic agents. This concept emerged from the hypothesis that these could protect the molecule of interest from undesirable interactions with components of the biological milieu along with improving its solubility. Several carriers have been studied: linear polymers, micellar assemblies, liposomes, polymersomes, and, more recently, dendritic structures (dendrimers and dendrons).<sup>(1)</sup> The ideal carrier should facilitate high drug loading, long blood circulation time (in the case of the commonly explored intravenous administration), high accumulation in the desired tissue, low toxicity, low immunogenicity, simplicity in its preparation, and, preferably, adequate biodegradability.

Dendritic structures emerged from a new class of highly branched polymers, first synthesized by Voegtli and colleagues in 1978, and coined "cascade molecules".<sup>(2)</sup> Later on, Denkewalter, Tomalia, Newkome, Frechet, and co-workers further increased the level of complexity of these branched molecules, giving rise to larger dendritic structures that were then renamed "dendrimers".<sup>(3)</sup> Dendrimers consist of the following: (a) a central core with two or more reactive groups, (b) repeated units or monomers covalently attached to the central core and organized in layers called "generations" (G), and (c) terminal functional groups on their surface. They can be synthesized by two different approaches: divergent<sup>(3b)</sup> or convergent.<sup>(3d)</sup>

The most commonly used dendrimers are poly(amido amine) (PAMAM),<sup>(3b)</sup> poly(propylene imine) (PPI),<sup>(2)</sup> and poly(lysine)-based dendrimers;<sup>(4)</sup> currently, all are commercially available. These and other dendrimers have been proposed as promising carriers for drug delivery<sup>(5)</sup> due to their unique structural characteristics: globular, well-defined, and very branched structure, as well as their monodispersion and controllable nanosize. Moreover, the presence of a high density of terminal functional groups allows the tethering of different ligands and/or drugs in a specific and controllable manner, simulating the multivalency present in different biological systems. This multivalency is the greater virtue of dendrimers: the enhanced effect that stems from presenting lots of several bioactive molecules at the same time and place. Additionally, dendrimers can also cargo a molecule of interest by forming nanosized structures stabilized by noncovalent interactions.

The characteristics of the surface groups of the dendrimers, besides determining predominantly their physicochemical properties, will also determine their biological activity and biocompatibility. For example, cationic dendrimers will more readily interact with the negatively charged surface of the cell membranes, but have also been found to have more cytotoxicity than anionic or neutral ones.<sup>(6)</sup> A common approach for masking the surface charge and, in general, improving the biocompatibility of dendrimers and/or dendrons, as well as increasing their circulation time in the bloodstream, is to tether to the dendrimers backbone chains of poly(ethylene glycol) (PEG).<sup>(7)</sup> This strategy was also carried out in the case of DEP docetaxel, a dendrimer with docetaxel attached that is in Phase 1 clinical trials for the treatment of a wide range of solid tumors including breast, lung, and prostate. Other dendrimers have also reached clinical trials: VivaGel, a G<sub>4</sub> poly(l-lysine)-based dendrimer, which acts as an antimicrobial agent being applied in the treatment/prevention of a range of sexually transmitted diseases, is in Phase 3 trials; and Gadomer-17, a poly(lysine)-based dendrimer bearing 24 Gd(III)-DOTA chelates (commercial name Dotarem; DOTA = 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid), is in Phase 2 trials for its use as a dendritic contrast agent for diagnosis in magnetic resonance imaging (MRI).

Despite the progress in the design of dendritic structures with improved features for biomedical application, one of the main drawbacks of the most currently used dendrimers is their nondegradability under physiological conditions that can result in cytotoxicity induced by the accumulation of nondegradable synthetic materials inside cells or in tissues.(8) In vitro studies have shown that dendrimer cytotoxicity is mainly associated with cell membrane disruption and subsequent necrosis/non-apoptotic cell death.(6a, 8d) Numerous works have thoroughly described the effect of dendrimer chemistry, size, and charge on biological membrane integrity.(8c, 9) However, recent reports suggest that, apart from membrane destabilization, toxicity may also arise from impaired oxidative metabolism resulting from mitochondrial dysfunction(10) and changes in endogenous gene expression(11) that ultimately lead to apoptotic cell death.(10, 11) Dendrimer chemistry, charge, and size are features that will also have an impact on in vivo biodistribution and pharmacokinetics.(6a, 8c, 9a) Additionally, in vivo cytotoxicity will also depend on the dose and the administration route.(6a) However, conflicting data can be found in the open literature concerning in vivo testing. Some studies report that PAMAM and PPI dendrimers, especially at low generations, are not as toxic as initially described.(12) However, others have reported toxicity profiles for the same dendrimers.(13) Consequently, a number of teams has been focusing on the design of biodegradable dendritic structures, to overcome these hurdles. The use of biodegradable materials that, under biological environmental conditions, degrade in time into smaller fragments that can be excreted or eliminated through metabolic pathways is expected to overcome the risk of complications associated with the long-term presence of high-molecular-weight compounds.

The development of biodegradable dendritic structures has also been put forward in the context of the design of “smart” controlled delivery systems in which one aims at triggering and/or sustaining the release of a therapeutic agent via the control of the degradation profile of its vector. Inclusively, some authors have proposed the use of degradable dendritic structures as contrasting agents that can overcome long-term Gd(III) tissue accumulation.(14)

Here, we present the state-of-the-art of biodegradable dendritic structures, with particular focus on the ones used in the context of drug and nucleic acid delivery, as well as their use as macromolecular diagnostic agents. The design and synthesis of such structures are presented, as well as the challenges that remain ahead toward their widespread application. Starting from the assumption that at least some animal research is relevant, ethically acceptable and presently not replaceable, some harm to animals in research may be perceived as a ‘necessary evil’, in particular in face of the moral importance of advancing biomedical knowledge for the benefit of humans and non-humans alike. However, it should nevertheless be reflected upon in which circumstances it may – or may not – be either ‘necessary’ or ‘evil’ to kill animals in the context of animal research. In this chapter, we discuss whether killing is inevitable, or morally problematic, as well as to whom this killing matters.

## 2. GETTING AND TUNING DEGRADABILITY

As in the case of linear polymers, the biodegradability of dendrimers can be achieved by inclusion in their structure of labile bonds to be broken due to a specific biological activity or stimulus. To the best of our knowledge, the majority of efforts have been focused on the development of dendritic architectures (dendrimers, dendrons, and linear–dendritic block copolymers) with hydrolyzable bonds. The functionalities more susceptible to hydrolytic cleavage are based on anhydrides, esters, phosphoesters, carbamates, ethers, and amides. The main factors that control the rate of degradation of dendritic structures are (a) the chemical bond present on or connecting the monomer units; (b) the hydrophobicity of the monomer units—more hydrophilic monomers result in faster degrading

structures compared to hydrophobic monomers; (c) size—larger dendrimers degrade more slowly compared to smaller ones due to the higher packaging; and (d) the localization of the cleavable linkages, since the hydrolysis of interior linkages leads to faster degradation of the whole dendrimer. With these parameters taken into account, it is expected that one can modulate the properties of biodegradable dendritic structures to achieve the desired degradation rate for a specific application.

The covalent degradation or dendrimer fragmentation could proceed through removal of a certain percentage of surface functionalities, removal of dendritic branches or monomers, or removal of the central core. Therefore, the cleavage of only certain parts of the backbone of the designed dendrimer could lead to its full degradation into low-molecular-weight fragments. Moreover, it is worthwhile mentioning that the degradation of some cleavage polymeric materials (like aliphatic polyesters) could also be accelerated by the presence of superoxide ions ( $O_2^-$ ) generated in the body fluids and tissues during inflammatory response to foreign materials.<sup>(15)</sup>

### 3. BIODEGRADABLE DENDRITIC STRUCTURES

The different types of biodegradable dendritic structures can be classified according to the type of degradable connection or bond, the monomer on which they are based, and the synthetic strategy to prepare them, among others.

Although very few examples of biodegradable dendritic structures not based on ester bonds can be found (see section 3.4), the majority of the dendrimers that are susceptible to degradation under physiological conditions reported so far contain ester bonds as focal points of degradation. Polyester dendrimers represent an attractive class of nanomaterials due to their good biocompatibility and the compromise between their biodegradability trait and the possibility of synthetic manipulation in comparison with other more hydrolytic susceptible structures, such as polyanhydrides. Even so, the synthesis of these nanocarriers is challenging because of the hydrolytic susceptibility of the ester bond.<sup>(16)</sup> In contrast, polyethers and polyamide-based dendrimers could withstand much wider selection of synthetic manipulations, yet they do not degrade easily in the body but only under severe conditions of hydrolysis. Thus, these dendritic structures may be more prone to long-term accumulation in vivo. Therefore, these types of dendrimers will not be considered in the context of this Review.

Aromatic polyester dendrimers were the first polyester dendrimers to be described in 1992, by Hawker and Fréchet.<sup>(17)</sup> With this work, they introduced the convergent synthetic route for the synthesis of dendrimers. Despite containing ester linkages, they are aromatic esters, with high hydrolytic stability. Thus, these structures can barely be regarded as biodegradable dendrimers. In 1996 the first study mentioning biodegradable dendrimers was reported, where the enzymatic degradation of aliphatic chiral polyester dendrimers based on (R)-3-hydroxybutanoic acid and trimesic acid was described (Figure 1).<sup>(18)</sup> Since then, several other polyester dendritic structures have been proposed: 2,2-bis(hydroxymethyl)propanoic acid (bis-HMPA)-derivatives (section 3.1), other aliphatic esters derivatives (section 3.2), and alternating polyesters (section 3.3). In this work, all the aliphatic polyester dendrimers will be considered biodegradable, even in those cases in which authors did not study their degradation profile.

In the following subsections, the design and synthesis of several biodegradable dendritic structures are presented.

### 3.1. Monomers Based on bis-HMPA.

Since its first use, 2,2-bis(hydroxymethyl)propanoic acid (bis-HMPA) (Figure 2a) has been the main monomer of choice in the synthesis of biodegradable dendritic structures. Bis-HMPA is commercially available at low cost, and the resulting polyester dendrimers are non-immunogenic, nontoxic, and biodegradable:(19) all very attractive features when envisaging biological applications.

The first report on the synthesis of polyester dendrimers based on bis-HMPA was by Ihre, Hult, and Söderlind (Figure 2c).(20) Since then, a considerable number of research groups proposed several alternative synthetic routes and different precursors using different protecting groups (Figure 2b), with the aim of improving and simplifying the synthesis of this family of biodegradable dendrimers until higher generations with good yields. Additionally, different functionalizations of the core and/or on the surface were reported by several authors to use these dendrimers for different purposes.(21)

In 2002, Gillies and Fréchet described dendrimers consisting of two bis-HMPA dendrons covalently attached as a “bow-tie” (“bow-tie dendrimers”, Figure 2d) aiming at water-soluble drug carriers.(22)

In 2004, Vestberg et al. reported the synthesis of bis-HMPA dendrons up to G<sub>5</sub> with porphyrins as core.(23) Porphyrins were used because of their potential applications in many areas, such as photodynamic therapy, nanosensors, and so forth.(24) Bis-HMPA dendrimers containing carboranes, which interest resides in their potential use in boron neutron capture therapy in cancer treatment and will be revisited in more detail in section 4, were synthesized by Parrott(21m) and Galie.(25) Gillies and Fréchet reported the synthesis of polyester dendritic bow-ties with a bis(adamantylurea)-glycinyurea system at the focal point of the bow-tie.(26) They prepared self-assembled polyester bow-tie dendrimers with various peripheral groups.

Bis-HMPA polyester dendrimers presenting cyclic carbonates on their periphery were also reported. The attraction of this approach is that carbonate groups react with amines, even in water, yielding bifunctional products easily.(27)

Some studies that combined identical dendrons via the Diels–Alder reaction to furnish symmetrical dendrimers were reported.(28) In 2008, Kose et al. reported the first example of the synthesis of segment block dendrimers using the Diels–Alder-based synthetic strategy in order to get unsymmetrical dendrimers.(29) The cycloaddition is very attractive in dendrimer synthesis, since no reagents are required. Therefore, the resulting dendrimers are free of impurities such as toxic metals, which represent a problem for biological applications.

Several authors reported linear–dendritic block copolymers for different purposes including drug delivery, where the characteristics of biodegradable bis-HMPA dendrons are combined with linear polymers, also biodegradable in some cases. Polyester dendronized polymers (linear polymers having polyester dendrons at each repeating unit) that are expected to present an extended conformation were also reported by Grayson and Fréchet (based on a nondegradable linear polymer: poly(p-hydroxystyrene))(30) and Lee et al. (based on a degradable linear polymer: poly(ε-caprolactone) (PCL)).(31) Due to their high molecular weight and multivalency, these systems are also considered as promising vectors for drug delivery applications. The degradation half-life at 37 °C for the dendronized hydrophilic PCL was 2.5 days at pH 9.0 and 16 days at pH 7.4 (physiological conditions), while no changes in polymer molecular weight was observed under acidic conditions (pH 5.0, endolysosomal conditions).(31) These authors determined that, in fact, the dominant mode of degradation of these

ester bonds was under mildly basic conditions (pH 9.0). Additionally, the dendronized hydrophilic polymer was found to degrade much faster than the parent hydrophobic polymer.(32)

Connal et al. explored the known “arm first” synthetic strategy to synthesize cross-linked star polymers, with bis-HMPA dendrons at the end of the arms. Bis-HMPA dendrons up to the G<sub>5</sub> were synthesized and functionalized at the focal point with a single alkyl halide, capable of initiating polymerization of styrene by atom transfer radical polymerization (ATRP) (Figure 2f).(33).

In 2009, a series of well-defined and amphiphilic dumbbell-shaped triblock copolymers consisting of comb-shaped poly(L-lactide) (PLLA) end arms and linear PEG center block connected using bis-HMPA dendrons, with narrow molecular weight distributions and varied PLLA arm lengths, were presented (Figure 2e).(34) These dendronized linear polymers provide an alternative dendritic architecture with a variety of aspect ratios accessible depending on the length of the PLLA arms and PEG block, and the generation of the dendrons. Efficient regulation of material properties and cell responses was achieved using this series of compounds, that suggests their potential for various regenerative medicine and tissue-engineering applications.(34) Subsequently, a series of well-defined dumbbell-shaped triblock copolymers consisting of linear PEG linked to the focal point of bis-HMPA dendrons and comb-like poly( $\epsilon$ -caprolactone) with different arm lengths were further used to prepare microspheres with potential application as carriers for controlled delivery of water-soluble drugs.(35)

Recently, diblock and triblock dendron-polymer conjugates containing biodegradable bis-HMPA polyester dendron blocks and PEG were synthesized using the Diels-Alder “click” cycloaddition reaction by Gok et al.(36)

Kempe et al. reported the synthesis, in a one-pot cascade reaction approach, of a bis-HMPA dendron functionalized at the focal point with poly(2-ethyl-2-oxazoline) (Pox). Pox is a polymer with similar properties to PEG, such as protein repellency and stealth behavior; thus, it is being extensively discussed as a potential alternative to the use of PEG.(37)

Recently, Feliu et al. carried out a comprehensive assessment of the in vitro biocompatibility and degradability of a library of hydroxyl terminated bis-HMPA dendrimers of five different generations and their corresponding dendrons, that revealed excellent biocompatibility for these materials in the model systems used.(38) Besides the expected pH dependence of the degradation rate, in line with the previously reported data for bis-HMPA dendronized PCL,(31) it was found that the hydrolysis occurs first at the dendrimer periphery and progresses toward the core, through a mechanism of depolymerization. Additionally, sterically crowded G<sub>4</sub>-OH dendrimers were found to be extensively degraded after 2 days of incubation at pH 7.5.(38)

Thus, despite the widespread use of these bis-HMPA-based dendritic structures, some issues regarding premature or undesired degradation still remain to be addressed. Namely, the unwanted degradation of the PEGylated polyester backbone observed during the attachment of certain drugs.(39) Also, a significant degradation after only 10 h under physiological conditions (pH 7.4 and 37 °C) was observed by van der Poll et al. for a G<sub>1</sub> bis-HMPA-core dendrimer, which limits their potential use in certain biological applications.(16a) In order to solve this problem, these authors reported a synthesis that combines the biocompatibility and biodegradability of polyester dendrimers with the robustness of polyamide dendrimers, to yield a hybrid scaffold capable of translation into in vivo studies. In this manner, the degradation rate was significantly reduced, with the degradation profile for these ester-amide hybrid dendrimers being prolonged up to 20 days.(16a).



### 3.2 Other Aliphatic Ester Monomers

Other aliphatic ester monomers have been used to synthesize polyester dendrimers, although to a smaller extent than bis-HMPA. Carnahan and Grinstaff have reported polyester dendrimers coined as "biodendrimers", which are composed of natural metabolites and biocompatible with the human body. These authors developed several dendrimers based on only succinic acid, only adipic acid, and mixtures of the two (Figure 3a).<sup>(40)</sup> The properties of the mixed dendrimers depends on the composition of the outer generation layer.<sup>(40c)</sup> The esterification of the dendrimers with succinic acid monomethylallyl ether, and subsequent photochemical polymerization of the alkenes, yielded soft gels.<sup>(40a)</sup> These materials have been explored as corneal adhesives<sup>(41)</sup> and for cartilage repair.<sup>(42)</sup>

The synthesis of another family of biodegradable cationic dendrimers was proposed in 2006 (Figure 3b).<sup>(43)</sup> The ester-amine dendrons and dendrimers synthesized in this work by the convergent method up to G<sub>3</sub> and G<sub>2</sub>, respectively, were based on bis(2-hydroxy-ethyl)-amino]-acetic acid tert-butyl ester as the growing unit (Figure 3b), with internal tertiary amines and protected terminal amines. The synthesized dendrons and dendrimers, with their terminal amines deprotected, were degradable in D<sub>2</sub>O at 37 °C. The degradation rates were found to be dependent on the generation number and the ability of the primary amines to access and to catalyze the ester group degradation. All dendrons were completely degraded within 30 days, while only 60% of the sterically crowded dendrimers were degraded over the same time period.<sup>(43)</sup>

Twibanire et al. synthesized tribranched dendrons in order to prepare polyester dendrimers with denser layers than those derived from bis-HMPA and, therefore, more resistant under physiological conditions.<sup>(21g)</sup> The synthesis of polyester dendrimers using as starting materials benzyl acetoacetate and tert-butyl acrylate (Figure 3c) were presented by Hirayama et al.<sup>(44)</sup> Similar dendrimers, also designed to be applied as drug delivery vectors, were prepared from 3-hydroxyacetophenone and its tert-butyldimethylsilyl ether.<sup>(45)</sup>

Polyester dendrimers containing tertiary amines were prepared by Bouillon et al.<sup>(46)</sup> The amine groups can serve as buffers to neutralize the protons delivered from the ester hydrolysis; therefore, these poly(ester amine) dendrimers are very attractive as drug delivery vectors (Figure 3d).

Another interesting contribution was the synthesis of polyester dendrimers presenting functional groups capable of orthogonal reactions,<sup>(47)</sup> rendering bifunctional dendrimers.<sup>(48)</sup> An AB<sub>2</sub>C dendron (Figure 3e.1) was used to originate dendrimers bearing alkyne units and hydroxyl groups (Figure 3e.2). A bifunctional dendrimer hosting azide and alcohol functionalities was also synthesized.<sup>(48)</sup>

In 2013, Akiyama et al. presented divergent and convergent procedures for the synthesis of another type of poly(ester amine) dendrimers, having an adamantane structure as core, which allows that even low generation dendrimers have a globular structure.<sup>(49)</sup> Recently, Pahovnik and et al. also reported on the synthesis of different generation, water-soluble poly(ester amide) dendrimers with hydroxyl functional groups from an AB<sub>2</sub> adduct of bis-HMPA and glycine as repeating unit and 1,1,1-tris(hydroxymethyl)propane as core. These poly(ester amide) dendrimers were used for solubilizing an anti-diabetic drug.<sup>(50)</sup>

### 3.3 Alternating Polyester Dendrimers

Different types of dendrimers, called by some authors “alternating dendrimers”,<sup>(51)</sup> can be produced in which the ester linkages alternate with other types of linkages, if orthogonal coupling methods are used in alternation.<sup>(48, 51, 52)</sup> The alternation of click reactions with ester formation using two different AB<sub>2</sub> dendrons (Figure 4a.1) for the accelerated synthesis of the dendrimers was reported by Antoni et al.<sup>(53)</sup> Because these orthogonal reactions do not require protecting groups and, thus, activation or deprotection steps are not needed, only five steps led to the preparation of a G<sub>4</sub> dendrimer (Figure 4a.2).

Ma et al. developed a facile synthesis of biodegradable alternating polyester dendrimers from sequential click coupling of asymmetrical monomers (2-[(methacryloyl)oxy]ethyl acrylate (MAEA) and cysteamine), with pendant methacrylate or amine groups that can be easily used for conjugations of drugs.<sup>(54)</sup> Later, Ye et al. simplified the synthesis by using a  $\beta$ -cyclodextrin core, from which polyester dendrimers with high molecular weights could be easily synthesized in fewer steps without intensive purifications.<sup>(14)</sup> Moreover, they carried out the *in vivo* evaluation of these biodegradable dendrimers as MRI contrast agents, the data of which will be further discussed in section 6.<sup>(14)</sup> Montañez et al. used AB<sub>2</sub> dendrons for the development of another approach for the accelerated synthesis of polyester dendrimers with terminal alkene bonds.<sup>(55)</sup> Walter et al. developed a series of macrothiols bearing latent hydroxyls (Figure 4b.1).<sup>(56)</sup> Dendrimers based on bis-HMPA were obtained through the photochemically induced addition of these macrothiols to alkene-terminated core molecules. Deprotection of the latent hydroxyls originated a hydroxyl-terminated dendrimer (Figure 4b.2) that can then be functionalized further to give products with desired properties.<sup>(56)</sup>

An efficient convergent synthetic route alternating esterification with light-promoted addition of a thiol to an alkyne was described by Chen et al. This dendrimer was shown to effectively bind the anticancer drug cis-dichlorodiammineplatinum(II).<sup>(57)</sup> Rosen et al. presented another alternating synthetic polyester dendrimers route for synthesizing dendrimers containing  $\alpha$ -bromo esters, which can polymerize by a single electron transfer living radical polymerization and thus produce star polymers with low generations of dendrimer units at the center and on the periphery.<sup>(58)</sup> Downing et al. reported that acyloxysilyl bonds (silyl esters) can be incorporated into dendritic structures as an easily degradable bond.<sup>(59)</sup>

In 2013, Khoei et al. published the preparation of pH-susceptible linear–dendritic–linear block copolymers from poly( $\epsilon$ -caprolactone), asymmetric poly(ester amine) dendrons (G<sub>1</sub>–G<sub>3</sub>), and PEG.<sup>(60)</sup> Their hydrolytic degradation was investigated at two pHs (5.8 and 7.4) at 37 °C. They observed that the copolymers with the higher-generation dendrons degraded more slowly than those with lower generations. Furthermore, in this case, the degradation rate at pH 5.8 is faster than at pH 7.4 for all generations. These amphiphilic systems can self-assemble and form micelles in water. Thus, they designed nanoparticles containing magnetite coated with these block copolymers for encapsulating hydrophobic drugs, such as quercetin. Their sensitivity at lower pHs is an attractive feature for using these nanoparticles for drug delivery *in vivo*.<sup>(60)</sup>

### 3.4 Degradable Dendrimers Not Based on Ester Bonds

An approach to obtain polyacetal systems using sequential transacetalation and protection–deprotection techniques has been proposed by Fuchs and Lemcoff for the preparation of macromolecular polyacetals that were subsequently applied to obtain new dendrimers, which may undergo hydrolysis to effectively “unzip” the dendrimer to polyfunctional macromolecules or degrade it altogether.<sup>(61)</sup>



Using an iterative and divergent approach, Zimmerman and co-workers synthesized dendrimers from degradable 1,3,5-triazaadamantanes (TAA) monomers at each branch point (Figure 4c).(62) Contrary to some polyester dendrimers, TAAs are stable under basic and physiological conditions (pH 7.4), but hydrolyze rapidly (half-life lower than 30 min) under acidic conditions (e.g., endosomal pH) to give basic and well-defined byproducts.(62a)

In the following sections, the applications of biodegradable dendritic structures as vectors of both drugs and nucleic acids, as well as their function as macromolecular contrast agents in MRI, are revisited.

#### 4 Biodegradable Dendritic Structures in Drug Delivery

One of the first applications of dendrimers in the biomedical field was as drug delivery vectors, since they can easily transport drug molecules in their interior and/or tethered to their terminal surface groups.(5a, 63)

The advantages of using dendrimers as drug carriers are multiple and include the following: (a) the possibility of being designed to carry hydrophobic or hydrophilic drugs, protecting them from degradative processes or unwanted interactions with biological molecules; (b) the capacity to simultaneously release two or more drugs at the site of interest allowing the tuning of pharmacodynamics; (c) the potential to modify the dendrimer surface to confer hydrophilicity to the dendrimers, to help them to surpass important biological barriers (such as those imposed by the mononuclear phagocyte system or cellular membranes), or to endow them with chemical ligands that are recognized by cell receptors and actively target specific types of cells or tissues; (d) the chance to play with the dendrimer size (generation) to modulate the dendrimer's blood circulating time and/or passively target solid tumors through the enhanced permeability and retention (EPR) effect; (e) the ability to achieve a controlled delivery of the drugs by controlling the dendrimer/drug interactions or by developing stimuli-responsive systems; and (f) the possibility to use dendrimers as theranostic platforms through their combination with chemical entities/nanoparticles suitable as contrast agents in bioimaging (this point will be further explored in section 6).(5a, 63)

Biodegradable dendrimers show all the mentioned potentialities of dendrimers for drug delivery, further presenting the advantage of being transformed under physiological conditions into small-size products that may be metabolized or excreted from the body. In this context, the dendrimers and their products of degradation should show a low level of toxicity, as is the case for polyester dendrimers.(19, 38, 64) About 12 years ago, in a representative work, Padilla De Jesús evaluated the toxicity both in vitro and in vivo of several dendritic structures based on bis-HMPA.(64) All the structures were shown to have a low cytotoxicity and a high tolerability upon administration in mice by intravenous (i.v.) bolus injection. One of these structures, a high-molecular-weight 3-arm PEG-dendrimer hybrid (that exhibited the longest circulatory half-life) was conjugated to doxorubicin by means of an acid-labile hydrazone linkage. In vitro experiments showed that the system was able to release the drug as a function of pH with a higher release at pH below 6. Results also indicated that the cytotoxicity of the drug was significantly reduced after linking it to the dendrimer, demonstrating that the use of a carrier alters the pharmacokinetics and the distribution of the drug. The design of the dendrimer structure to prevent rapid and premature degradation was also revealed as an important feature toward obtaining an effective drug delivery system. Later, the same group reported a study where orthogonal "bow-tie" polyester dendrimers were prepared with PEG arms connected to one of the polyester dendrons via degradable carbamate bonds (Figure 2d).(19) Here, the polyester dendron was conjugated with the

drug and, while complete ester hydrolysis alone should release the bis-HMPA monomers, carbamate hydrolysis should release the individual PEG arms. A library of eight polyester dendrimer-PEG bow-tie hybrids was prepared (molecular weights from 20 000 to 160 000, and number of PEG arms ranging from two to eight) and evaluated as controlled drug release systems. In this work, as expected, the high-molecular-weight carriers (>40 000) exhibited longer circulation half-lives. Furthermore, the in vitro degradability of the G<sub>3</sub> bow-tie with 10 kDa PEG was followed for 15 days at pH 9.0, pH 7.4, and pH 5.0 and 37 °C. The reported results were in line with the ones reported by Lee et al. for the bis-HMPA dendronized PCL.<sup>(31)</sup> Additionally, the obtained results showed that at pH 5.0 only the carbamate bonds degrade, while at pH 7.4 the ester bond present in the bis-HMPA dendron backbone also degrades.<sup>(19)</sup> A subsequent study showed the effectiveness of these carriers as drug vehicles.<sup>(65)</sup> Indeed, a single injection of a high-molecular-weight dendrimer-PEG-doxorubicin conjugate substantially inhibited the progression of a doxorubicin-insensitive C-26 colon carcinoma upon i.v. administration to BALB/c mice. A PEGylated dendrimer based on a polyester-polyamide hybrid backbone was also described for doxorubicin transport through conjugation.<sup>(16a)</sup> As previously mentioned at the end of section 3.1, the design of the carrier was chosen to diminish destructive side reactions during dendrimer synthesis and premature degradation, while a suitable biodegradability is equally maintained. The anticancer efficiency of the dendrimer-doxorubicin conjugate was compared to that of Doxil using equal dosages in the treatment of C26 murine colon carcinoma. In this work, statistically equivalent results were obtained with the two systems with most mice tumor-free at the end of the two-month experiment, with the test group animals indicating that the ester-amide dendrimer may exhibit less toxicity than Doxil at equivalent doxorubicin dosages.<sup>(16a)</sup>

The encapsulation of drugs inside dendritic structures is a common strategy in drug delivery that can help in the solubilization of hydrophobic drugs such as the camptothecins that have low water solubility but high anticancer efficiency. Morgan et al. used one of their “biodendrimers”, the one based on glycerol and succinic acid, for the encapsulation of 10-hydroxycamptothecin and 7-butyl-10-aminocamptothecin.<sup>(66)</sup> The cytotoxicity of the dendrimer-drug complexes toward four different human cancer cell lines was analyzed showing low IC<sub>50</sub> values. Importantly, cellular uptake and drug retention were increased in MCF-7 cells when using the prepared dendrimer.<sup>(66)</sup> In another study, Dhanikula and Hildgen investigated the influence of the architecture of polyester-co-polyether (PEPE) dendrimers on the encapsulation and release of the anticancer drug methotrexate.<sup>(67)</sup> In vitro experiments showed that the different dendrimers were biocompatible and presented a good capacity to encapsulate this drug. An increase in the number of branches and in the size of internal cavities was shown to enhance the encapsulation capacity, while the absence of aromatic rings as branching units substantially decreased the loading capacity of PEPE dendrimers. The authors concluded that the mechanisms of encapsulation involved physical entrapment, weak hydrogen bonding, and hydrophobic interactions.<sup>(67)</sup>

Interestingly, thermoreversible hydrogels were prepared by Namazi and Adeli from dendronized polymers based on citric acid (dendron part) and PEG (linear part).<sup>(68)</sup> It should be highlighted that several hydrophobic anti-inflammatory drugs have been successfully encapsulated inside the formed gels. Lundberg et al. presented bis-HMPA dendrons of different generations (G<sub>0</sub>–G<sub>4</sub>) as functionalized macroinitiators for the construction of sophisticated amphiphilic dendritic-linear hybrid materials with hydrophobic poly( $\epsilon$ -caprolactone) at the chain-ends and a single strain of hydrophilic PEG at the core.<sup>(69)</sup> These were utilized for the fabrication of drug loaded micelles as well as the development of isoporous membranes.

Though still at a preliminary stage, “disassembled”, “cleavable”, or “self-immolative” dendrimers (SIDs) that respond to an external stimulus (e.g., pH variation, enzymatic action, catalytic antibodies, among others) constitute very promising systems for drug delivery.<sup>(70)</sup> These dendrimers have a unique structural degradable backbone, the most common cleavage functionalities of which are esters and carbamates, that allows a cascade decomposition upon a simple triggering event. If drug molecules are incorporated as the tail units in these dendrimers and a suitable stimulus is used as the trigger, a multi-prodrug is generated that may be activated by a single cleavage event. The advantages associated with these dendritic prodrugs are significant when compared with monomeric prodrugs in inhibition of tumor cell growth.<sup>(71)</sup>

Biodegradable dendrimers have also been used in the delivery of boron-10 for neutron capture therapy (NCT) of cancer. For this, it is crucial to ensure an adequate boron concentration at the target sites, and as such, high boron content species (like carboranes) can be used in the design of the carriers. The polyester dendrimers based on bis-HMPA proposed by Parrott et al. incorporated several carboranes within its structure (section 3.1).<sup>(21m)</sup> Fourth- and fifth-generation dendrimers that contained 4, 8, and 16 carborane cages were prepared. The irradiation of these dendrimers with thermal neutrons resulted in emission of  $\gamma$  radiation, thus showing the occurrence of boron neutron capture events and the potentiality of using these dendrimers in cancer treatment.<sup>(21m)</sup>

## 5 Biodegradable Dendritic Structures for Nucleic Acid Delivery

Over the past decades, gene therapy has emerged as a promising therapeutic approach to prevent and treat several diseases/conditions. Its underlying principles are the modulation of gene and/or protein expression of the host cells following the introduction of exogenous nucleic acids (DNA, RNA, or single-stranded oligonucleotides) into somatic cells.<sup>(72)</sup>

Nucleic acids (NAs) are susceptible to degradation by serum endonucleases, further decreasing their already low internalization when administered naked.<sup>(1e)</sup> Moreover, following cellular uptake, NAs are susceptible to additional degradation both in the lysosomal/endosomal pathway and in the cytoplasm.<sup>(72, 73)</sup> Thus, most of the gene therapy strategies proposed so far rely on delivery vectors that, ideally, should contribute to the overcoming of different extra- and intracellular barriers in order to efficiently deliver NAs into cells with minimal toxicity.<sup>(72, 73)</sup> Briefly, the main hurdles toward a successful gene therapeutic intervention include the following: (a) NA degradation by endonucleases present in the extracellular milieu, (b) cellular internalization, (c) endosomal escape, (d) NA release from the vector and access to the cytoplasmic or nuclear target, and (e) vector intra- and extracellular accumulation. Nuclear delivery, necessary in the case of DNA<sup>(72, 73)</sup> or certain oligonucleotides,<sup>(74)</sup> further requires that the NA crosses the nuclear membrane. Additionally, the delivery vector must avoid unspecific binding to serum proteins, preventing aggregation.<sup>(5c, 72a, 73)</sup> NA vectors are divided into two main categories: viral and nonviral vectors.<sup>(72a, 73)</sup> Despite the high transfection rates observed for viral vectors, insertional mutagenesis and immunogenicity together with both low scale production and carrying capacity are delaying their application as safe and viable gene therapy vectors.<sup>(72a, 73, 75)</sup> Consequently, these obstacles have drawn much attention to the development of nonviral vectors, such as lipids, polymers, and dendrimers.<sup>(1e, 72a, 73)</sup> A common feature among these has been their cationic nature. The same characteristics that make dendrimers attractive platforms for drug delivery are equally important when it comes to the delivery of therapeutic NAs. Their adaptable and tunable chemistry together with the high density of functional groups allows the design of an almost unlimited number of molecules and the conjugation of several ligands tailored to attain efficient

gene delivery.(5c, 63a) Nevertheless, much research is needed for these to progress into preclinical and clinical developments.

Cationic dendrimers are able to complex NAs through electrostatic interactions between their positively charged terminal groups and the negatively charged phosphate groups of the NA, originating dendriplexes.(5c, 72a) This process occurs in a concentration-dependent manner and is also affected by the medium properties, such as pH, temperature, and ionic concentration.(5c)

Even though several groups reported the success of dendrimers as NA delivery vehicles,(7b, 7h, 76) the nonbiodegradability of the dendritic structures used remains a drawback yet to be solved.(6a, 8d, 63a) As previously mentioned, the ideal gene delivery vehicle should be biodegradable to prevent bioaccumulation and subsequent cytotoxicity.(8) Moreover, biodegradability can contribute to the carrier's multivalency decreasing as a function of time, leading to a lower interaction with the transported NA, promoting its release.(77)

So far, the few reports in the open literature on the use of biodegradable dendritic structures for gene delivery have been based on the application of bis-HMPA-based dendrons. Welsh et al. synthesized bis-HMPA based dendrons up to the third generation (G<sub>3</sub>) with carbamate-linked spermine groups on their surface and a benzyl ester protecting group at the focal point.(77a) The efficiency of DNA binding increased in a generation dependent manner due to the enhanced multivalency. Degradation studies showed that the G<sub>3</sub> dendrimer was stable at pH 5.0, but degraded at the physiological pH 7.4 on the time scale of 8 days.(77a) In a slightly different approach, Barnard et al. replaced the benzyl ester protecting group in G<sub>2</sub> dendrons by hydrophobic units (cholesterol and hydrocarbon chains) to promote their controlled self-assembly rendering a system with higher multivalency, which significantly enhances the DNA binding. Furthermore, they modified the surface groups with N,N-di(3-aminopropyl)-N-(methyl)amine (DAPMA), a triamine that showed lower cytotoxicity than spermine.(77b) As expected, these dendrons remained intact at pH 5.0 and degraded at pH 7.5. Even though cholesterol-functionalized dendrons were shown to have lower degradation rates than the hydrocarbon-functionalized ones, every dendron was completely degraded after a period of 6–10 h.(77b) The reported G<sub>2</sub> dendrons could efficiently complex DNA and undergo cellular internalization, yet a low transfection efficiency was observed. The authors hypothesized that, at the lower endosomal pH, or when bound to DNA, the degradation of these dendrons becomes ineffective on the transfection time scale, which could explain their poor transfection performance.(77b) The same group further modified the cholesterol-functionalized G<sub>2</sub> dendrons with small PEG chains (triethylene glycol and octaethylene glycol).(78) PEG addition was found to affect size and zeta potential properties of the resulting dendriplexes and enhanced DNA binding to different extents, with a higher binding affinity observed for the structures based on the longer PEG chain.

More recently, Barnard and et al. have reported a different strategy aimed at improving DNA release and dendron degradation.(79) They synthesized self-assembly disulfide-linked dendron nanoparticles, where the cholesterol groups were attached to the DAPMA-terminated G<sub>2</sub> bis-HMPA dendrons by an S–S linkage. In vitro, upon addition of dithiothreitol, nanoparticles were shown to undergo disassembly in 1 h due to cleavage of the disulfide linkage, which consequently led to loss of self-assembled multivalent binding, triggering DNA release (Figure 5). After that, the remaining fragments undergo further degradation over longer time scales (24 h), which validates a controllable double-degradation process.(79) Disulfide linkers are widely used in the design of nonviral gene delivery vectors due to their enhanced degradability in the reductive environment of the cytoplasm.(72)

The success of gene therapy is highly dependent on the development of efficient and safe NA delivery vectors. Additionally, it is extremely important to develop new biodegradable and biocompatible vectors to prevent both bioaccumulation and cytotoxicity, and easy clinical translation. Such vectors could arise from newly developed dendritic agents, which allow an almost unlimited combination of structures and architectures together with the ability to multiconjugate several target-molecules.

## 6 Biodegradable Dendritic Structures as Magnetic Resonance Imaging Contrast Agents

Currently, medical imaging plays a major role in health care management as one of the most efficient techniques for early and accurate diagnosis and treatment follow-up, expectedly leading to more successful treatment. As a noninvasive technique, MRI allows high spatial resolution and both physiological and anatomical information without resorting to harmful ionizing radiation. However, both its low sensitivity and target-specificity remain significant limitations.(5d)

MRI is based on the interaction of protons (mostly water protons) with each other and their surroundings. Upon the application of strong magnetic fields, the magnetic moments of protons will orient themselves in the direction of the magnetic field.(80) After disruption of this magnetic alignment they will return to the previous state in a process described by two relaxation rates: the longitudinal and the transversal relaxations, characterized by the time constants  $T_1$  and  $T_2$ , respectively.(80, 81) The relaxation rates vary from tissue to tissue, which gives rise to the image contrast in MRI. However, the inherent differences in relaxation times among tissues are not always enough to generate an adequate contrast. Thus, it is rather common to use exogenous contrast agents (CA), which act by reducing  $T_1$  and  $T_2$  through interaction with the surrounding water protons.(5d, 80, 81) The ability to decrease  $T_1$  and  $T_2$  is measured by the  $r_1$  and  $r_2$  relaxivities, respectively ( $r_1 = 1/T_1$ ;  $r_2 = 1/T_2$ ). (80, 81)

Contrast agents can be classified by their biodistribution, magnetic properties (paramagnetic and superparamagnetic), and effect on the image contrast, which can be positive ( $T_1$  reduction) or negative ( $T_2$  reduction), increasing or decreasing the signal intensity, respectively.(82) Paramagnetic agents are predominantly  $T_1$ -reducing CA, while superparamagnetic usually originate  $T_2$  reductions.(82) The most widely used CAs are small molecules based on paramagnetic gadolinium(III) chelates, such as Gd(III)-DTPA (DTPA = diethylenetriaminepentaacetic acid, Magnevist) and Gd(III)-DOTA (Dotarem).(5d, 83) Administered prior to the MR imaging, these CAs are known for their contrast enhancement. Yet, their low molecular relaxivity, nonspecificity, and fast renal clearance require the usage of potentially harmful high doses, which clearly limits image quality.(5d) Conjugation of Gd(III)-chelates with polymers,(84) proteins,(85) dendrimers,(5d) or other scaffolds(86) can enhance relaxivity and blood circulation times improving image contrast.(1f)

Due to their multivalency, dendrimers represent one of the most appealing platforms to carry multiple CAs moieties.(5d) Moreover, their branched structure brings an extra rigidity and the possibility to graft target molecules, offering a unique opportunity to enhance site-specific image contrast (Figure 6).(81)

The applicability of dendrimers as MRI contrast agents (dendritic CAs, DCAs) was first demonstrated by Wiener et al. where a PAMAM dendrimer conjugated with a Gd(III)-DTPA showed an ionic relaxivity of  $34 \text{ mM}^{-1} \text{ s}^{-1}$ , meaning a 6-fold higher relaxivity compared to that of Gd(III)-DTPA alone ( $5.4 \text{ mM}^{-1} \text{ s}^{-1}$ ). (87) Additionally, they observed that these novel structures were able to carry more Gd(III) ions than other existing macromolecules at the time, which could possibly be increased at higher generations. Although the macromolecular CAs developed so far have been yielding good image

contrast and enhanced sensitivity, the large sizes decrease renal excretion, which in turn significantly increases the possibility of bioaccumulation and further release of toxic Gd(III) ions.(5d, 83, 88) In fact, some studies have reported that nephrogenic fibrosis and other severe diseases were associated with the use of certain Gd(III) chelates, particularly in patients with impaired renal function.(89) So far, only Gadomer-17, a poly(lysine)-based dendritic CA bearing 24 Gd(III)-DOTA chelates, has advanced into clinical evaluation due to its almost exclusive intravascular biodistribution and rapid renal clearance.(90)

Thus, recent efforts have been made to develop biocompatible and biodegradable DCAs (Table 1). These maintain the nondegradable DCAs characteristics, such as blood circulation times and enhanced image contrast, and at the same time can be easily degraded under physiological conditions into smaller byproducts preventing accumulation and subsequent toxicity.

Nazemi et al. designed a dendritic Gd(III)-DTPA functionalized polymersome.(91) The azide-terminated polymersome consisted of a PCL-PEG block copolymer and a G<sub>3</sub> bis-HMPA polyester (PE) dendron. Ionic relaxivities ( $r_1$ ) of 12.1 and 26.1 mM<sup>-1</sup> s<sup>-1</sup> (20 MHz and 25 °C) were observed for the dendron alone and the dendron-functionalized polymersomes, respectively.(91) While most of the DCAs developed so far have been targeted to  $r_1$  augmentation, Klemm et al. focused on designing contrast agents to improve  $r_2$ .(92, 94) T<sub>1</sub> CAs usually lose efficiency at higher magnetic fields, which constitutes an important limitation since clinical instruments work at increased magnetic fields to enhance image contrast. In contrast, T<sub>2</sub> CAs exhibit higher relaxivities at higher magnetic fields.(92, 94) As reported by Klemm et al., a versatile CA with the ability to enhance  $r_1$  and  $r_2$  would bring an excellent opportunity to improve both imaging modes.(92) In this sense, they conjugated a Gd(III)-TACN-bis(1-Me)-3,2-HOPO-TAM-ethylamine to an ester-amide (EA) dendrimer via an amide bond.(92) Remarkably, these complexes were able to increase Gd(III)  $r_1$  and  $r_2$  up to 31 and 52 mM<sup>-1</sup> s<sup>-1</sup>, respectively. This represents a 10-fold increase in  $r_1$  compared to that of the clinically available CAs. Moreover, this DCA was shown to be nontoxic at concentrations up to 25 μM at 72 h.(92) Other studies published by the same group described similar DCAs conjugated with other chelating molecules that enhanced  $r_1$  and  $r_2$ , as well.(94, 95) As mentioned in section 3.3, in 2012, Ma et al. synthesized G<sub>0</sub> to G<sub>3</sub> alternating polyester dendrimers, based on MAEA and cysteamine monomers with a β-cyclodextrin core.(54) This dendrimer was further conjugated with Gd(III)-DTPA and evaluated in vivo as DCA.(14) G<sub>0</sub> had a relaxivity of 10.6 mM<sup>-1</sup> s<sup>-1</sup>, while G<sub>1</sub>, G<sub>2</sub>, and G<sub>3</sub> showed similar relaxivity values around 11.7 mM<sup>-1</sup> s<sup>-1</sup>. This phenomenon was attributed to the comparable internal flexibility of the higher generations, which hindered the expected rise of the relaxivity with the generation. Again, degradation studies, using the G<sub>3</sub> dendrimer, showed that at mildly acidic pH little or no degradation was detected, while at pH 7.4 only 70% of the ester bonds remained intact 20 h post-incubation. This percentage decreased to 35% in the presence of an esterase. For in vivo studies the authors introduced zwitterions into the dendrimer to enhance solubility. The zwitterionic G<sub>2</sub> DCA was shown to have higher blood circulation times and image improvement than the clinically used Magnevist. Additionally, this dendrimer was shown to promote lower long-term Gd(III) accumulation in several tissues when compared to the nondegradable dendritic MRI agent PAMAM-G6-DO<sub>3</sub>A-Gd(III) (DO<sub>3</sub>A = 1,4,7-tricarboxymethyl-1,4,7,10-tetraazacyclododecane).(14)

More recently, molecular MRI has also drawn attention due to its potential use in the dissection of disease mechanisms and other events at the cellular and subcellular levels, which can be achieved by combining CAs with target molecules.(5d, 96) In 2012, the G<sub>2</sub> polyester dendrimer reported above was further functionalized with PEG and folic acid (FA) to target tumor cells overexpressing folic acid receptors.(14, 89a) PEGylation resulted in enhanced relaxivity due to the increasing solubility and



complex stiffness. FA-PEG-G<sub>2</sub>-DTPA-Gd(III) had a higher and longer tumor image contrast than that of Magnevist and a much lower Gd(III) accumulation compared with nondegradable contrast agents, due to dendrimer degradation. Degradation studies showed that 30% of the ester bonds of this DCA hydrolyzed at pH 7.4 against 7% at pH 5.0 after 10 h. The hydrolyzed ester bonds at pH 7.4 were shown to increase to 47% in the presence of an esterase.<sup>(89a)</sup> Despite the great performance of this DCA, the need for solubilizing groups might hamper translation to the clinic.<sup>(93)</sup> In order to overcome this, the same team developed a new DCA based on the same polyester dendrimer, but with a tris(2-aminoethyl)amine core to increase solubility without further modifications (PEGylation), hence preventing high molecular weights and size distribution.<sup>(93)</sup> As expected, they observed that  $r_1$  increased from 10.2 to 17.5 mM<sup>-1</sup> s<sup>-1</sup> when going from G<sub>1</sub> to G<sub>5</sub> dendrimer, demonstrating a generation-dependent relaxivity. Interestingly, in vivo MRI revealed major biodistribution differences between the G<sub>2</sub> and G<sub>5</sub> dendrimers, as the first enhanced tumor contrast whereas the second enhanced liver signal. The smaller size of the G<sub>2</sub> allowed improved tumor imaging due to the passive tumor targeting via the EPR effect.<sup>(93, 97)</sup> As expected, G<sub>2</sub>-DTPA-Gd(III) displayed pH-dependent degradability, being stable at pH 5.0 and degraded at pH 7.4 (20 h post-incubation, 80% of the ester bonds were degraded).<sup>(93)</sup>

Biodegradable dendritic-based MRI contrast agents have already proven to be a promising alternative to the currently available CAs. Even though the significant molecular weight increase could be a limitation to the clinical setting, it remains true that lower amounts of CA would be necessary due to the relaxivity increase. Therefore, further testing is needed to formulate an adequate molecular weight/relaxivity ratio. Success could rely on the optimization of already existing systems, as well as on the development of new biodegradable and biocompatible dendritic platforms. Moreover, the combination of several MRI labels and target-specific molecules in the same dendritic scaffold could give rise to better and more accurate diagnostics.

## 7 Conclusions and Future Perspectives

The use of degradable polymeric systems has a wide perspective in several established and emerging biomedical technologies, including controlled-release systems for targeted drug delivery, antibacterial drugs, and tissue regeneration. As the demand increases, a higher level of control over the structure, properties, and development of degradable materials is being pursued.

Dendrimers allow this coveted structural control. Among these, biodegradable dendrimers represent an attractive class of nanomaterials. They possess two major advantages over the conventional dendrimers, which confers an additional inherent “smartness” to these systems: (a) multiple covalently bound bioactive molecules can be site-specifically released from the targeted dendrimer by a single cleaving step, and (b) they are degraded and therefore can be easily eliminated from the body avoiding the toxicity related to the accumulation of synthetic materials in cells/tissues. Consequently, biodegradable dendrimers are expected to surpass the limitations of the most used nondegradable systems. Even so, the design and synthesis of biodegradable dendritic structures soluble in water with precise branching, molecular weight, monodispersion, and with a suitable pharmacodynamics continues to be a challenge. Other important key aspects are to overcome the undesired degradation of the degradable backbone observed during the attachment of certain functional groups and/or bioactives, as well as to limit the premature hydrolysis of the backbone and/or functional moieties in order to ensure their efficient application. In fact, these hurdles may explain the reduced number of reports where biodegradable dendrimers are applied for a particular function, in comparison with the higher number of works reporting only the design and synthesis of biodegradable dendrimers. This will

require considerable effort in the development of new synthetic routes for the development of these systems.

Nevertheless, once these drawbacks are surpassed, biodegradable dendrimers are expected to recast the existing therapeutic practices. The simultaneous combination of multivalency and biodegradability with precise architectures will definitely make dendrimers a greater platform, versatile for many biomedical applications as revised here.

## Acknowledgements

The authors would like to acknowledge the FEDER funds through the Programa Operacional Factores de Competitividade – COMPETE and the Portuguese funds through FCT – Fundação para a Ciência e a Tecnologia (PTDC/CTM-NAN/112428/2009, PEst/SAU/LA0002/2013 and PEst-OE/QUI/UI0674/2013-2014) that supported this work. V. Leiro is supported by FCT (SFRH/BPD/69110/2010).

## REFERENCES

- 1(a) Gregoriadis, G. (1988) Liposomes as drug carriers: Recent trends and progress, John Wiley and Sons, Chichester.
- (b) Torchilin, V. P. (1991) Immobilized Enzymes in Medicine, Springer-Verlag, Berlin.
- (c) Rolland, A. (1993) Pharmaceutical Particulate Carriers: Therapeutic Applications, Marcel Dekker, New York.
- (d) Sahoo, S. K., Jain, T. K., Reddy, M. K., and Labhasetwar, V. (2008) Nano-Sized Carriers for Drug Delivery, In NanoBioTechnology (Oded, S. and Levy, I., Eds.) pp 329–348, Humana Press, Totowa, NJ.
- (e) Mintzer, M. A. and Simanek, E. E. (2009) Nonviral Vectors for Gene Delivery Chem. Rev. 109, 259–302
- (f) Tang, J., Sheng, Y., Hu, H., and Shen, Y. (2013) Macromolecular MRI contrast agents: Structures, properties and applications Prog. Polym. Sci. 38, 462–502
- 2Buhleier, E., Wehner, W., and Vogtle, F. (1978) "Cascade"- and "non-skid-chain-like" syntheses of molecular cavity topologies Synthesis 2, 155–158
- 3(a) Denkwalter, R. G., Kolc, J., and Lukasavage, W. J. Surface modifying agents, metal chelating agents, substrates for drugs. U.S. Patent 4,289,872, 1981.
- (b) Tomalia, D. A., Baker, H., Dewald, J., Hall, M., Kallos, G., Martin, S., Roeck, J., Ryder, J., and Smith, P. (1985) A new class of polymers: starburst-dendritic macromolecules Polym. J. 17, 117–132
- (c) Newkome, G. R., Yao, Z. Q., Baker, G. R., and Gupta, V. K. (1985) Micelles. Part 1. Cascade molecules: a new approach to micelles. A [27]-arbo J. Org. Chem. 50, 2003–2004

(d) Hawker, C. J. and Frechet, J. M. J. (1990) Preparation of polymers with controlled molecular architecture. A new convergent approach to dendritic macromolecules *J. Am. Chem. Soc.* 112, 7638–7647

4(a) Rao, C. and Tam, J. P. (1994) Synthesis of peptide dendrimer *J. Am. Chem. Soc.* 116, 6975–6976

(b) Tam, J. P. (1988) Synthetic peptide vaccine design: synthesis and properties of a high-density multiple antigenic peptide system *Proc. Natl. Acad. Sci. U. S. A.* 85, 5409–5413

5(a) Svenson, S. and Tomalia, D. (2005) Dendrimers in biomedical applications—reflections on the field *Adv. Drug Delivery Rev.* 57, 2106–2129

(b) Medina, S. H. and El-Sayed, M. E. H. (2009) Dendrimers as carriers for delivery of chemotherapeutic agents *Chem. Rev.* 109, 3141–3157

(c) Dufes, C., Uchegbu, I., and Schatzlein, A. (2005) Dendrimers in gene delivery *Adv. Drug Delivery Rev.* 57, 2177–2202

(d) Langereis, S., Dirksen, A., Hackeng, T. M., van Genderen, M. H. P., and Meijer, E. W. (2007) Dendrimers and magnetic resonance imaging *New J. Chem.* 31, 1152–1160

(e) Sadler, K. and Tam, J. P. (2002) Peptide dendrimers: applications and synthesis *Rev. Mol. Biotechnol.* 90, 195–229

6(a) Duncan, R. and Izzo, L. (2005) Dendrimer biocompatibility and toxicity *Adv. Drug Delivery Rev.* 57, 2215–2237

(b) Stasko, N. A., Johnson, C. B., Schoenfisch, M. H., Johnson, T. A., and Holmuhamedov, E. L. (2007) Cytotoxicity of polypropylenimine dendrimer conjugates on cultured endothelial cells *Biomacromolecules* 8, 3853–3859

7(a) van Vlerken, L. E., Vyas, T. K., and Amiji, M. M. (2007) Poly(ethylene glycol)-modified nanocarriers for tumor-targeted and intracellular delivery *Pharm. Res.* 24, 1405–1414

(b) Luo, D., Haverstick, K., Belcheva, N., Han, E., and Saltzman, W. M. (2002) Poly(ethylene glycol)-conjugated PAMAM dendrimer for biocompatible, high-efficiency DNA delivery *Macromolecules* 35, 3456–3462

(c) Männistö, M., Vanderkerken, S., Toncheva, V., Elomaa, M., Ruponen, M., Schacht, E., and Urtti, A. (2002) Structure–activity relationships of poly(l-lysines): effects of pegylation and molecular shape on physicochemical and biological properties in gene delivery *J. Controlled Release* 83, 169–182

(d) Merdan, T., Kunath, K., Petersen, H., Bakowsky, U., Voigt, K. H., Kopecek, J., and Kissel, T. (2005) PEGylation of poly(ethylene imine) affects stability of complexes with plasmid DNA under in vivo conditions in a dose-dependent manner after intravenous injection into mice *Bioconjugate Chem.* 16, 785–792

- (e) Wood, K. C., Little, S. R., Langer, R., and Hammond, P. T. (2005) A family of hierarchically self-assembling linear-dendritic hybrid polymers for highly efficient targeted gene delivery *Angew. Chem., Int. Ed.* 44, 6704–6708
- (f) Kim, T.-i., Baek, J.-u., Yoon, J. K., Choi, J. S., Kim, K., and Park, J.-s. (2007) Synthesis and characterization of a novel arginine-grafted dendritic block copolymer for gene delivery and study of its cellular uptake pathway leading to transfection *Bioconjugate Chem.* 18, 309–317
- (g) Kim, T.-i., Seo, H. J., Choi, J. S., Jang, H.-S., Baek, J.-u., Kim, K., and Park, J.-S. (2004) PAMAM-PEG-PAMAM: novel triblock copolymer as a biocompatible and efficient gene delivery carrier *Biomacromolecules* 5, 2487–2492
- (h) de la Fuente, M., Raviña, M., Sousa-Herves, A., Correa, J., Riguera, R., Fernandez-Megia, E., Sánchez, A., and Alonso, M. J. (2012) Exploring the efficiency of gallic acid-based dendrimers and their block copolymers with PEG as gene carriers *Nanomedicine* 7, 1667–1681
- 8(a) Barth, R. F., Adams, D. M., Soloway, A. H., Alam, F., and Darby, M. W. (1994) Boronated starburst dendrimer-monoclonal antibody immunoconjugates: evaluation as a potential delivery system for neutron capture therapy *Bioconjugate Chem.* 5, 58–66
- (b) Malik, N., Evagorou, E., and Duncan, R. (1999) Dendrimer-platinate: a novel approach to cancer chemotherapy *Anticancer Drugs* 10, 767–776
- (c) Malik, N., Wiwattanapatapee, R., Klopsch, R., Lorenz, K., Frey, H., Weener, J. W., Meijer, E. W., Paulus, W., and Duncan, R. (2000) Dendrimers: relationship between structure and biocompatibility in vitro, and preliminary studies on the biodistribution of <sup>125</sup>I-labelled polyamidoamine dendrimers in vivo *J. Controlled Release* 65, 133–148
- (d) Jain, K., Kesharwani, P., Gupta, U., and Jain, N. K. (2010) Dendrimer toxicity: Let's meet the challenge *Int. J. Pharm.* 394, 122–142
- 9(a) Roberts, J. C., Bhalgat, M. K., and Zera, R. T. (1996) Preliminary biological evaluation of polyamidoamine (PAMAM) Starburst dendrimers *J. Biomed. Mater. Res.* 30, 53–65
- (b) Zhang, Z. Y. and Smith, B. D. (2000) High-generation polycationic dendrimers are unusually effective at disrupting anionic vesicles: membrane bending model *Bioconjugate Chem.* 11, 805–814
- (c) Mecke, A., Uppuluri, S., Sassanella, T. M., Lee, D. K., Ramamoorthy, A., Baker, J. R., Jr., Orr, B. G., and Banaszak Holl, M. M. (2004) Direct observation of lipid bilayer disruption by poly(amidoamine) dendrimers *Chem. Phys. Lipids* 132, 3–14
- (d) Hong, S., Bielinska, A. U., Mecke, A., Keszler, B., Beals, J. L., Shi, X., Balogh, L., Orr, B. G., Baker, J. R., Jr., and Banaszak Holl, M. M. (2004) Interaction of poly(amidoamine) dendrimers with supported lipid bilayers and cells: hole formation and the relation to transport *Bioconjugate Chem.* 15, 774–782
- (e) Agashe, H. B., Dutta, T., Garg, M., and Jain, N. K. (2006) Investigations on the toxicological profile of functionalized fifth-generation poly (propylene imine) dendrimer *J. Pharm. Pharmacol.* 58, 1491–1498

- 10(a) Kuo, J. H. S., Jan, M. S., and Lin, Y. L. (2007) Interactions between U-937 human macrophages and poly(propyleneimine) dendrimers *J. Controlled Release* 120, 51– 59
- (b) Lee, J. H., Cha, K. E., Kim, M. S., Hong, H. W., Chung, D. J., Ryu, G., and Myung, H. (2009) Nanosized polyamidoamine (PAMAM) dendrimer-induced apoptosis mediated by mitochondrial dysfunction *Toxicol. Lett.* 190, 202– 207
- (c) Thomas, T. P., Majoros, I., Kotlyar, A., Mullen, D., Holl, M. M., and Baker, J. R., Jr. (2009) Cationic poly(amidoamine) dendrimer induces lysosomal apoptotic pathway at therapeutically relevant concentrations *Biomacromolecules* 10, 3207– 3214
- (d) Mukherjee, S. P., Lyng, F. M., Garcia, A., Davoren, M., and Byrne, H. J. (2010) Mechanistic studies of in vitro cytotoxicity of poly(amidoamine) dendrimers in mammalian cells *Toxicol. Appl. Pharmacol.* 248, 259– 268
- (e) Naha, P. C., Davoren, M., Lyng, F. M., and Byrne, H. J. (2010) Reactive oxygen species (ROS) induced cytokine production and cytotoxicity of PAMAM dendrimers in J774A.1 cells *Toxicol. Appl. Pharmacol.* 246, 91– 99
- (f) Mukherjee, S. P. and Byrne, H. J. (2013) Polyamidoamine dendrimer nanoparticle cytotoxicity, oxidative stress, caspase activation and inflammatory response: experimental observation and numerical simulation *Nanomedicine* 9, 202– 211
- (g) Naha, P. C. and Byrne, H. J. (2013) Generation of intracellular reactive oxygen species and genotoxicity effect to exposure of nanosized polyamidoamine (PAMAM) dendrimers in PLHC-1 cells in vitro *Aquat. Toxicol.* 132–133, 61– 72
- 11 Omid, Y., Hollins, A. J., Drayton, R. M., and Akhtar, S. (2005) Polypropyleneimine dendrimer-induced gene expression changes: the effect of complexation with DNA, dendrimer generation and cell type *J. Drug Targeting* 13, 431– 443
- 12(a) Okuda, T., Kawakami, S., Maeie, T., Niidome, T., Yamashita, F., and Hashida, M. (2006) Biodistribution characteristics of amino acid dendrimers and their PEGylated derivatives after intravenous administration *J. Controlled Release* 114, 69– 77
- (b) Chauhan, A. S., Diwan, P. V., Jain, N. K., and Tomalia, D. A. (2009) Unexpected in vivo anti-inflammatory activity observed for simple, surface functionalized poly(amidoamine) dendrimers *Biomacromolecules* 10, 1195– 1202
- (c) Chauhan, A. S., Jain, N. K., and Diwan, P. V. (2010) Pre-clinical and behavioural toxicity profile of PAMAM dendrimers in mice *Proc. R. Soc. London, Ser. A: Math. Phys.* 466, 1535– 1550
- (d) Ziemba, B., Janaszewska, A., Ciepluch, K., Krotewicz, M., Fogel, W. A., Appelhans, D., Voit, B., Bryszewska, M., and Klajnert, B. (2011) In vivo toxicity of poly(propyleneimine) dendrimers *J. Biomed. Mater. Res., Part A* 99, 261– 268

(e) Albertazzi, L., Gherardini, L., Brondi, M., Sato, S. S., Bifone, A., Pizzorusso, T., Ratto, G. M., and Bardi, G. (2013) In vivo distribution and toxicity of PAMAM dendrimers in the central nervous system depend on their surface chemistry *Mol. Pharmaceutics* 10, 249–260

13(a) Labieniec, M., Ulicna, O., Vancova, O., Glowacki, R., Sebekova, K., Bald, E., Gabryelak, T., and Watala, C. (2008) PAMAM G<sub>4</sub> dendrimers lower high glucose but do not improve reduced survival in diabetic rats *Int. J. Pharm.* 364, 142–149

(b) Li, C., Liu, H., Sun, Y., Wang, H., Guo, F., Rao, S., Deng, J., Zhang, Y., Miao, Y., Guo, C., Meng, J., Chen, X., Li, L., Li, D., Xu, H., Wang, H., Li, B., and Jiang, C. (2009) PAMAM nanoparticles promote acute lung injury by inducing autophagic cell death through the Akt-TSC2-mTOR signaling pathway *J. Mol. Cell. Biol.* 1, 37–45

(c) Jones, C. F., Campbell, R. A., Brooks, A. E., Assemi, S., Tadjiki, S., Thiagarajan, G., Mulcock, C., Weyrich, A. S., Brooks, B. D., Ghandehari, H., and Grainger, D. W. (2012) Cationic PAMAM dendrimers aggressively initiate blood clot formation *ACS Nano* 6, 9900–9910

14 Ye, M., Qian, Y., Shen, Y., Hu, H., Sui, M., and Tang, J. (2012) Facile synthesis and in vivo evaluation of biodegradable dendritic MRI contrast agents *J. Mater. Chem.* 22, 14369–14377

15 Lee, K. H. and Chu, C. C. (2000) The role of superoxide ions in the degradation of synthetic absorbable sutures *J. Biomed. Mater. Res.* 49, 25–35

16(a) van der Poll, D. G., Kieler-Ferguson, H. M., Floyd, W. C., Guillaudeu, S. J., Jerger, K., Szoka, F. C., and Fréchet, J. M. (2010) Design, synthesis, and biological evaluation of a robust, biodegradable dendrimer Bioconjugate Chem. 21, 764–773

(b) Szuromi, P. D. (2003) Triggering polymer destruction *Science* 302, 1863

17 Hawker, C. J. and Fréchet, J. M. J. (1992) Monodispersed dendritic polyesters with removable chain ends: a versatile approach to globular macromolecules with chemically reversible polarities *J. Chem. Soc., Perkin Trans. 1* 19, 2459–2469

18 Seebach, D., Herrmann, G. F., Lengweiler, U. D., Bachmann, B. M., and Amrein, W. (1996) Synthesis and enzymatic degradation of dendrimers from (R)-3-hydroxybutanoic acid and trimesic acid *Angew. Chem., Int. Ed. Engl.* 35, 2795–2797

19 Gillies, E. R., Dy, E., Fréchet, J. M. J., and Szoka, F. C. (2005) Biological evaluation of polyester dendrimer: poly(ethylene oxide) “bow-tie” hybrids with tunable molecular weight and architecture *Mol. Pharmaceutics* 2, 129–138

20 Ihre, H., Hult, A., and Söderlind, E. (1996) Synthesis, characterization, and <sup>1</sup>H NMR self-diffusion studies of dendritic aliphatic polyesters based on 2,2-bis(hydroxymethyl)propionic acid and 1,1,1-tris(hydroxyphenyl)ethane *J. Am. Chem. Soc.* 118, 6388–6395

21(a) Ihre, H., Hult, A., Fréchet, J. M. J., and Gitsov, I. (1998) Double-stage convergent approach for the synthesis of functionalized dendritic aliphatic polyesters based on 2,2-bis(hydroxymethyl)propionic acid *Macromolecules* 31, 4061–4068



- (b) Greenwald, R. B., Conover, C. D., and Choe, Y. H. (2000) Poly(ethylene glycol) conjugated drugs and prodrugs: a comprehensive review *Crit. Rev. Ther. Drug Carrier Syst.* 17, 101– 161
- (c) Ihre, H. R., Padilla De Jesús, O. L., Szoka, F. C., and Fréchet, J. M. J. (2002) Polyester dendritic systems for drug delivery applications: design, synthesis, and characterization *Bioconjugate Chem.* 13, 443– 452
- (d) Ihre, H., Padilla De Jesús, O. L., and Fréchet, J. M. J. (2001) Fast and convenient divergent synthesis of aliphatic ester dendrimers by anhydride coupling *J. Am. Chem. Soc.* 123, 5908– 5917
- (e) Malkoch, M., Malmström, E., and Hult, A. (2002) Rapid and efficient synthesis of aliphatic ester dendrons and dendrimers *Macromolecules* 35, 8307– 8314
- (f) Parrott, M. C., Benhabbour, S. R., Saab, C., Lemon, J. A., Parker, S., Valliant, J. F., and Adronov, A. (2009) Synthesis, radiolabeling, and bio-imaging of high-generation polyester dendrimers *J. Am. Chem. Soc.* 131, 2906– 2916
- (g) Twibanire, J.-d. A. K., Al-Mughaid, H., and Grindley, T. B. (2010) Synthesis of new cores and their use in the preparation of polyester dendrimers *Tetrahedron* 66, 9602– 9609
- (h) Hedrick, J. L., Trollsås, M., Hawker, C. J., Atthoff, B., Claesson, H., Heise, A., Miller, R. D., Mecerreyes, D., Jérôme, R., and Dubois, P. (1998) Dendrimer-like star block and amphiphilic copolymers by combination of ring opening and atom transfer radical polymerization *Macromolecules* 31, 8691– 8705
- (i) Annby, U., Malmberg, M., Pettersson, B., and Rehnberg, N. (1998) Benzylidene protected bis-MPA a convenient dendrimer building block *Tetrahedron Lett.* 39, 3217– 3220
- (j) Trollsås, M., Atthoff, B., Claesson, H., and Hedrick, J. L. (1998) Hyperbranched poly( $\epsilon$ -caprolactone)s *Macromolecules* 31, 3439– 3445
- (k) Hao, X., Nilsson, C., Jesberger, M., Stenzel, M. H., Malmström, E., Davis, T. P., Östmark, E., and Barner-Kowollik, C. (2004) Dendrimers as scaffolds for multifunctional reversible addition–fragmentation chain transfer agents: Syntheses and polymerization *J. Polym. Sci., Part A: Polym. Chem.* 42, 5877– 5890
- (l) Wang, L., Meng, Z., Yu, Y., Meng, Q., and Chen, D. (2008) Synthesis of hybrid linear-dendritic block copolymers with carboxylic functional groups for the biomimetic mineralization of calcium carbonate *Polymer* 49, 1199– 1210
- (m) Parrott, M. C., Marchington, E. B., Valliant, J. F., and Adronov, A. (2005) Synthesis and properties of carborane-functionalized aliphatic polyester dendrimers *J. Am. Chem. Soc.* 127, 12081– 12089
- 22 Gillies, E. R. and Fréchet, J. M. J. (2002) Designing macromolecules for therapeutic applications: polyester dendrimer/poly(ethylene oxide) “bow-tie” hybrids with tunable molecular weight and architecture *J. Am. Chem. Soc.* 124, 14137– 14146

23Vestberg, R., Nyström, A., Lindgren, M., Malmström, E., and Hult, A. (2004) Porphyrin-cored 2,2-bis(methylol)propionic acid dendrimers *Chem. Mater.* 16, 2794–2804

24(a) Armstrong, N. R. (2000) Phthalocyanines and porphyrins as materials *J. Porphyrins Phthalocyanines* 4, 414–417

(b) Nishiyama, N., Stapert, H. R., Zhang, G.-D., Takasu, D., Jiang, D.-L., Nagano, T., Aida, T., and Kataoka, K. (2003) Light-harvesting ionic dendrimer porphyrins as new photosensitizers for photodynamic therapy *Bioconjugate Chem.* 14, 58–66

(c) Krivokapic, A., Anderson, H. L., Bourhill, G., Ives, R., Clark, S., and McEwan, K. J. (2001) Meso-tetra-alkynyl porphyrins for optical limiting—a survey of group III and IV metal complexes *Adv. Mater.* 13, 652–656

25Galie, K. M., Mollard, A., and Zharov, I. (2006) Polyester-based carborane-containing dendrons *Inorg. Chem.* 45, 7815–7820

26Gillies, E. R. and Fréchet, J. M. J. (2004) Synthesis and self-assembly of supramolecular dendritic “bow-ties”: effect of peripheral functionality on association constants *J. Org. Chem.* 69, 46–53

27Goodwin, A. P., Lam, S. S., and Fréchet, J. M. J. (2007) Rapid, efficient synthesis of heterobifunctional biodegradable dendrimers *J. Am. Chem. Soc.* 129, 6994–6995

28(a) McElhanon, J. R. and Wheeler, D. R. (2001) Thermally responsive dendrons and dendrimers based on reversible furan-maleimide Diels–Alder adducts *Org. Lett.* 3, 2681–2683

(b) Szalai, M. L., McGrath, D. V., Wheeler, D. R., Zifer, T., and McElhanon, J. R. (2007) Dendrimers based on thermally reversible furan-maleimide Diels–Alder adducts *Macromolecules* 40, 818–823

29Kose, M. M., Yesilbag, G., and Sanyal, A. (2008) Segment block dendrimers via Diels–Alder cycloaddition *Org. Lett.* 10, 2353–2356

30Grayson, S. M. and Fréchet, J. M. J. (2001) Divergent synthesis of dendronized poly(p-hydroxystyrene) *Macromolecules* 34, 6542–6544

31Lee, C. C., Grayson, S. M., and Fréchet, J. M. J. (2004) Synthesis of narrow-polydispersity degradable dendronized aliphatic polyesters *J. Polym. Sci., Part A: Polym. Chem.* 42, 3563–3578

32Ye, W. P., Du, F. S., Jin, J. Y., Yang, J. Y., and Xu, Y. (1997) In vitro degradation of poly(caprolactone), poly(lactide) and their block copolymers: Influence of composition, temperature and morphology *React. Funct. Polym.* 32, 161–168

33Connal, L. A., Vestberg, R., Hawker, C. J., and Qiao, G. G. (2007) Synthesis of dendron functionalized core cross-linked star polymers *Macromolecules* 40, 7855–7863

34Gong, F., Cheng, X., Wang, S., Wang, Y., Gao, Y., and Cheng, S. (2009) Biodegradable comb-dendritic tri-block copolymers consisting of poly(ethylene glycol) and poly(l-lactide): Synthesis, characterizations, and regulation of surface morphology and cell responses *Polymer* 50, 2775–2785

35Ju, M., Shen, L., Gong, F., Gao, Y., and Zhang, W. (2012) Synthesis and characterization of new biodegradable comb-dendritic triblock copolymers Polym. Int. 61, 1447–1455

36Gok, O., Yigit, S., Kose, M. M., Sanyal, R., and Sanyal, A. (2013) Dendron–polymer conjugates via the Diels–Alder “click” reaction of novel anthracene-based dendrons J. Polym. Sci., Part A: Polym. Chem. 51, 3191–3201

37Kempe, K., Onbulak, S., Schubert, U. S., Sanyal, A., and Hoogenboom, R. (2013) pH degradable dendron-functionalized poly(2-ethyl-2-oxazoline) prepared by a cascade “double-click” reaction Polym. Chem. 4, 3236–3244

38Feliu, N., Walter, M. V., Montañez, M. I., Kunzmann, A., Hult, A., Nyström, A., Malkoch, M., and Fadeel, B. (2012) Stability and biocompatibility of a library of polyester dendrimers in comparison to polyamidoamine dendrimers Biomaterials 33, 1970–1981

39Guillaudeu, S. J., Fox, M. E., Haidar, Y. M., Dy, E. E., Szoka, F. C., and Fréchet, J. M. J. (2008) PEGylated dendrimers with core functionality for biological applications Bioconjugate Chem. 19, 461–469

40(a) Carnahan, M. A. and Grinstaff, M. W. (2001) Synthesis and characterization of poly(glycerol–succinic acid) dendrimers Macromolecules 34, 7648–7655

(b) Grinstaff, M. W. (2002) Biodendrimers: new polymeric biomaterials for tissue engineering Chem.—Eur. J. 8, 2838–2846

(c) Carnahan, M. A. and Grinstaff, M. W. (2006) Synthesis of generational polyester dendrimers derived from glycerol and succinic or adipic acid Macromolecules 39, 609–616

41(a) Oelker, A. M. and Grinstaff, M. W. (2008) Ophthalmic adhesives: a materials chemistry perspective J. Mater. Chem. 18, 2521–2536

(b) Berdahl, J. P., Johnson, C. S., Proia, A. D., Grinstaff, M. W., and Kim, T. (2009) Comparison of sutures and dendritic polymer adhesives for corneal laceration repair in an in vivo chicken model Arch. Ophthalmol. 127, 442–447

42Söntjens, S. H. M., Nettles, D. L., Carnahan, M. A., Setton, L. A., and Grinstaff, M. W. (2006) Biodendrimer-based hydrogel scaffolds for cartilage tissue repair Biomacromolecules 7, 310–316

43Lee, J. S., Huh, J., Ahn, C. H., Lee, M., and Park, T. G. (2006) Synthesis of novel biodegradable cationic dendrimers Macromol. Rapid Commun. 27, 1608–1614

44Hirayama, Y., Nakamura, T., Uehara, S., Sakamoto, Y., Yamaguchi, K., Sei, Y., and Iwamura, M. (2005) Synthesis and characterization of polyester dendrimers from acetoacetate and acrylate Org. Lett. 7, 525–528

45Hirayama, Y., Sakamoto, Y., Yamaguchi, K., Sakamoto, S., and Iwamura, M. (2005) Synthesis of polyester dendrimers and dendrons starting from Michael reaction of acrylates with 3-hydroxyacetophenone Tetrahedron Lett. 46, 1027–1030

46Bouillon, C., Tintaru, A., Monnier, V., Charles, L., Quéléver, G., and Peng, L. (2010) Synthesis of poly(amino)ester dendrimers via active cyanomethyl ester intermediates *J. Org. Chem.* 75, 8685–8688

47The use of orthogonal coupling reactions is one powerful strategy that consists of using compatible functional groups for avoiding protecting groups. Avoiding protection/deprotection steps can dramatically streamline the construction of complex molecules, such as dendrimers.

48Antoni, P., Hed, Y., Nordberg, A., Nyström, D., von Holst, H., Hult, A., and Malkoch, M. (2009) Bifunctional dendrimers: from robust synthesis and accelerated one-pot postfunctionalization strategy to potential applications *Angew. Chem., Int. Ed.* 48, 2126–2130

49Akiyama, H., Miyashita, K., Hari, Y., Obika, S., and Imanishi, T. (2013) Synthesis of novel polyesteramine dendrimers by divergent and convergent methods *Tetrahedron* 69, 6810–6820

50Pahovnik, D., Čusak, A., Reven, S., and Žagar, E. (2014) Synthesis of poly(ester-amide) dendrimers based on 2,2-Bis(hydroxymethyl) propanoic acid and glycine *J. Polym. Sci., Part A: Polym. Chem.* 52, 3292–3301

51Twibanire, J.-d. A. K. and Grindley, T. B. (2012) Polyester dendrimers *Polymers* 4, 794–879

52Zeng, F. and Zimmerman, S. C. (1996) Rapid synthesis of dendrimers by an orthogonal coupling strategy *J. Am. Chem. Soc.* 118, 5326–5327

53Antoni, P., Nyström, D., Hawker, C. J., Hult, A., and Malkoch, M. (2007) A chemoselective approach for the accelerated synthesis of well-defined dendritic architectures *Chem. Commun.* 2249–2251

54Ma, X., Tang, J., Shen, Y., Fan, M., Tang, H., and Radosz, M. (2009) Facile synthesis of polyester dendrimers from sequential click coupling of asymmetrical monomers *J. Am. Chem. Soc.* 131, 14795–14803

55Montañez, M. I., Campos, L. M., Antoni, P., Hed, Y., Walter, M. V., Krull, B. T., Khan, A., Hult, A., Hawker, C. J., and Malkoch, M. (2010) Accelerated growth of dendrimers via thiol-ene and esterification reactions *Macromolecules* 43, 6004–6013

56Walter, M. V., Lundberg, P., Hult, A., and Malkoch, M. (2011) Novel macrothiols for the synthesis of a structurally comprehensive dendritic library using thiol-ene click chemistry *J. Polym. Sci., Part A: Polym. Chem.* 49, 2990–2995

57Chen, G., Kumar, J., Gregory, A., and Stenzel, M. H. (2009) Efficient synthesis of dendrimers via a thiol-yne and esterification process and their potential application in the delivery of platinum anti-cancer drugs *Chem. Commun.* 6291–6293

58(a) Rosen, B. M., Lligadas, G., Hahn, C., and Percec, V. (2009) Synthesis of dendrimers through divergent iterative thio-bromo “click” chemistry *J. Polym. Sci., Part A: Polym. Chem.* 47, 3931–3939

(b) Rosen, B. M., Lligadas, G., Hahn, C., and Percec, V. (2009) Synthesis of dendritic macromolecules through divergent iterative thio-bromo “click” chemistry and SET-LRP *J. Polym. Sci., Part A: Polym. Chem.* 47, 3940–3948

59Downing, C. M., Missaghi, M. N., Kung, M. C., and Kung, H. H. (2011) Design and synthesis of readily degradable acyloxysilane dendrimers *Tetrahedron* 67, 7502–7509

60Khoee, S. and Hemati, K. (2013) Synthesis of magnetite/polyamino-ester dendrimer based on PCL/PEG amphiphilic copolymers via convergent approach for targeted diagnosis and therapy *Polymer* 5574–5585

61Lemcoff, N. G. and Fuchs, B. (2002) Toward novel polyacetals by transacetalation techniques: dendrimeric diacetals *Org. Lett.* 4, 731–734

62(a) Balija, A. M., Kohman, R. E., and Zimmerman, S. C. (2008) Substituted 1,3,5-triazaadamantanes: biocompatible and degradable building blocks *Angew. Chem., Int. Ed.* 47, 8072–8074

(b) Kohman, R. E. and Zimmerman, S. C. (2009) Degradable dendrimers divergently synthesized via click chemistry *Chem. Commun.* 794–796

63(a) Mintzer, M. A. and Grinstaff, M. W. (2010) Biomedical applications of dendrimers: a tutorial *Chem. Soc. Rev.* 40, 173–190 [PubMed],

(b) Gillies, E. R. and Fréchet, J. M. J. (2005) Dendrimers and dendritic polymers in drug delivery *Drug Discovery Today* 10, 35–43

(c) Menjoge, A. R., Kannan, R. M., and Tomalia, D. A. (2010) Dendrimer-based drug and imaging conjugates: design considerations for nanomedical applications *Drug Discovery Today* 15, 171–185

(d) Astruc, D., Boisselier, E., and Ornelas, C. (2010) Dendrimers designed for functions: from physical, photophysical, and supramolecular properties to applications in sensing, catalysis, molecular electronics, photonics, and nanomedicine *Chem. Rev.* 110, 1857–1959

64Padilla De Jesús, O. L., Ihre, H. R., Gagne, L., Fréchet, J. M. J., and Szoka, F. C. (2002) Polyester dendritic systems for drug delivery applications: in vitro and in vivo evaluation *Bioconjugate Chem.* 13, 453–461

65Lee, C. C., Gillies, E. R., Fox, M. E., Guillaudeau, S. J., Fréchet, J. M. J., Dy, E. E., and Szoka, F. C. (2006) A single dose of doxorubicin-functionalized bow-tie dendrimer cures mice bearing C-26 colon carcinomas *Proc. Natl. Acad. Sci. U. S. A.* 103, 16649–16654

66Morgan, M. T., Nakanishi, Y., Kroll, D. J., Griset, A. P., Carnahan, M. A., Wathier, M., Oberlies, N. H., Manikumar, G., Wani, M. C., and Grinstaff, M. W. (2006) Dendrimer-encapsulated camptothecins: increased solubility, cellular uptake, and cellular retention affords enhanced anticancer activity in vitro *Cancer Res.* 66, 11913–11921

67Dhanikula, R. S. and Hildgen, P. (2007) Influence of molecular architecture of polyether-co-polyester dendrimers on the encapsulation and release of methotrexate *Biomaterials* 28, 3140–3152

68(a) Namazi, H. and Adeli, M. (2003) Novel linear–globular thermoreversible hydrogel ABA type copolymers from dendritic citric acid as the A blocks and poly(ethyleneglycol) as the B block *Eur. Polym. J.* 39, 1491–1500

(b) Namazi, H. and Adeli, M. (2005) Dendrimers of citric acid and poly (ethylene glycol) as the new drug-delivery agents *Biomaterials* 26, 1175– 1183

69Lundberg, P., Walter, M. V., Montañez, M. I., Hult, D., Hult, A., Nyström, A., and Malkoch, M. (2010) Linear dendritic polymeric amphiphiles with intrinsic biocompatibility: synthesis and characterization to fabrication of micelles and honeycomb membranes *Polym. Chem.* 2, 394– 402

70(a) Wang, R. E., Costanza, F., Niu, Y., Wu, H., Hu, Y., Hang, W., Sun, Y., and Cai, J. (2012) Development of self-immolative dendrimers for drug delivery and sensing *J. Controlled Release* 159, 154– 163

(b) Amir, R. J., Danieli, E., and Shabat, D. (2007) Receiver–amplifier, self-immolative dendritic device *Chem.—Eur. J.* 13, 812– 821

(c) Amir, R. J. and Shabat, D. (2004) Self-immolative dendrimer biodegradability by multi-enzymatic triggering *Chem. Commun.* 1614– 1615

71(a) Haba, K., Popkov, M., Shamis, M., Lerner, R. A., Barbas, C. F., and Shabat, D. (2005) Single-triggered trimeric prodrugs *Angew. Chem., Int. Ed.* 44, 716– 720

(b) Shamis, M., Lode, H. N., and Shabat, D. (2004) Bioactivation of self-immolative dendritic prodrugs by catalytic antibody 38C2 *J. Am. Chem. Soc.* 126, 1726– 1731

72(a) Jones, C. H., Chen, C.-K., Ravikrishnan, A., Rane, S., and Pfeifer, B. A. (2013) Overcoming nonviral gene delivery barriers: perspective and future *Mol. Pharmaceutics* 10, 4082– 4098

(b) Gomes, C. P., Lopes, C. D. F., Moreno, P. M. D., Varela-Moreira, A., Alonso, M. J., and Pêgo, A. P. (2014) Translating chitosan to clinical delivery of nucleic acid-based drugs *MRS Bull.* 39, 60– 70

73Nishikawa, M. and Huang, L. (2001) Nonviral vectors in the new millennium: delivery barriers in gene transfer *Hum. Gene Ther.* 12, 861– 870

74Kole, R., Krainer, A. R., and Altman, S. (2012) RNA therapeutics: beyond RNA interference and antisense oligonucleotides *Nat. Rev. Drug Discovery* 11, 125– 140

75Clare, E. T., Anja, E., and Mark, A. K. (2003) Progress and problems with the use of viral vectors for gene therapy *Nat. Rev. Genet.* 4, 346– 358 [PubMed],

76(a) Raviña, M., de la Fuente, M., Correa, J., Sousa-Herves, A., Pinto, J., Fernandez-Megia, E., Riguera, R., Sanchez, A., and Alonso, M. J. (2010) Core–shell dendriplexes with sterically induced stoichiometry for gene delivery *Macromolecules* 43, 6953– 6961

(b) Nouri, A., Castro, R., Kairys, V., Santos, J. L., Rodrigues, J., Li, Y., and Tomás, H. (2012) Insight into the role of N,N-dimethylaminoethyl methacrylate (DMAEMA) conjugation onto poly(ethylenimine): cell viability and gene transfection studies *J. Mater. Sci. Mater. Med.* 23, 2967– 2980



- (c) Pandita, D., Santos, J. L., Rodrigues, J., Pêgo, A. P., Granja, P. L., and Tomás, H. (2011) Gene delivery into mesenchymal stem cells: a biomimetic approach using RGD nanoclusters based on poly(amidoamine) dendrimers *Biomacromolecules* 12, 472–481
- (d) Rodrigues, J., Jardim, M. G., Figueira, J., Gouveia, M., Tomás, H., and Rissanen, K. (2011) Poly(alkylidenamines) dendrimers as scaffolds for the preparation of low-generation ruthenium based metallodendrimers *New J. Chem.* 35, 1938–1943
- (e) Santos, J. L., Oliveira, H., Pandita, D., Rodrigues, J., Pêgo, A. P., Granja, P. L., and Tomás, H. (2010) Functionalization of poly(amidoamine) dendrimers with hydrophobic chains for improved gene delivery in mesenchymal stem cells *J. Controlled Release* 144, 55–64
- (f) Santos, J. L., Oramas, E., Pêgo, A. P., Granja, P. L., and Tomás, H. (2009) Osteogenic differentiation of mesenchymal stem cells using PAMAM dendrimers as gene delivery vectors *J. Controlled Release* 134, 141–148
- (g) Santos, J. L., Pandita, D., Rodrigues, J., Pêgo, A. P., Granja, P. L., Balian, G., and Tomás, H. (2010) Receptor-mediated gene delivery using PAMAM dendrimers conjugated with peptides recognized by mesenchymal stem cells *Mol. Pharmaceutics* 7, 763–774
- 77(a) Welsh, D. J., Jones, S. P., and Smith, D. K. (2009) “On-off” multivalent recognition: degradable dendrons for temporary high-affinity DNA binding *Angew. Chem., Int. Ed.* 48, 4047–4051
- (b) Barnard, A., Posocco, P., Prich, S., Calderon, M., Haag, R., Hwang, M. E., Shum, V. W., Pack, D. W., and Smith, D. K. (2011) Degradable self-assembling dendrons for gene delivery: experimental and theoretical insights into the barriers to cellular uptake *J. Am. Chem. Soc.* 133, 20288–20300
- 78Barnard, A., Calderon, M., Tschiche, A., Haag, R., and Smith, D. K. (2012) Effects of a PEG additive on the biomolecular interactions of self-assembled dendron nanostructures *Org. Biomol. Chem.* 10, 8403–8409
- 79Barnard, A., Posocco, P., Fermeglia, M., Tschiche, A., Calderon, M., Prich, S., and Smith, D. K. (2014) Double-degradable responsive self-assembled multivalent arrays – temporary nanoscale recognition between dendrons and DNA *Org. Biomol. Chem.* 12, 446–455
- 80Villaraza, A. J. L., Bumb, A., and Brechbiel, M. W. (2010) Macromolecules, dendrimers, and nanomaterials in magnetic resonance imaging: the interplay between size, function, and pharmacokinetics *Chem. Rev.* 110, 2921–2959
- 81Bumb, A., Brechbiel, M. W., and Choyke, P. (2010) Macromolecular and dendrimer-based magnetic resonance contrast agents *Acta Radiol.* 51, 751–767
- 82Caravan, P. (2006) Strategies for increasing the sensitivity of gadolinium based MRI contrast agents *Chem. Soc. Rev.* 35, 512–523
- 83Langereis, S., Lussanet, Q. G. d., van Genderen, M. H. P., Backes, W. H., and Meijer, E. W. (2004) Multivalent contrast agents based on gadolinium–diethylenetriaminepentaacetic acid-terminated poly(propylene imine) dendrimers for magnetic resonance imaging *Macromolecules* 37, 3084–3091

84(a) Duarte, M. G., Gil, M. H., Peters, J. A., Colet, J. M., Elst, L. V., Muller, R. N., and Geraldes, C. F. G. C. (2001) Synthesis, characterization, and relaxivity of two linear Gd(DTPA)-polymer conjugates Bioconjugate Chem. 12, 170–177

(b) Zhang, W. L., Yong, D. W., Huang, J., Yu, J. H., Liu, S. Y., and Fan, M. X. (2011) Fabrication of polymer-gadolinium (III) complex nanomicelle from poly(ethylene glycol)-polysuccinimide conjugate and diethylenetriaminetetraacetic acid-gadolinium as magnetic resonance imaging contrast agents J. Appl. Polym. Sci. 120, 2596–2605

85Liepold, L., Anderson, S., Willits, D., Oltrogge, L., Frank, J. A., Douglas, T., and Young, M. (2007) Viral capsids as MRI contrast agents Magn. Reson. Med. 58, 871–879

86Lin, W.-I., Lin, C.-Y., Lin, Y.-S., Wu, S.-H., Huang, Y.-R., Hung, Y., Chang, C., and Mou, C.-Y. (2012) High payload Gd(III) encapsulated in hollow silica nanospheres for high resolution magnetic resonance imaging J. Mater. Chem. B 1, 639–645

87Wiener, E., Brechbiel, M. W., Brothers, H., Magin, R. L., Gansow, O. A., Tomalia, D. A., and Lauterbur, P. C. (1994) Dendrimer-based metal chelates: A new class of magnetic resonance imaging contrast agents Magn. Reson. Med. 31, 1–8

88Venditto, V. J., Regino, C. A. S., and Brechbiel, M. W. (2005) PAMAM dendrimer based macromolecules as improved contrast agents Mol. Pharmaceutics 2, 302–311

89(a) Ye, M., Qian, Y., Tang, J., Hu, H., Sui, M., and Shen, Y. (2013) Targeted biodegradable dendritic MRI contrast agent for enhanced tumor imaging J. Controlled Release 169, 239–245

(b) Thomsen, H. S. (2009) Nephrogenic systemic fibrosis: history and epidemiology Radiol. Clin. North Am. 47, 827–829

(c) Bhawan, J., Swick, B. L., Koff, A. B., and Stone, M. S. (2009) Sclerotic bodies in nephrogenic systemic fibrosis: a new histopathologic finding J. Cutaneous Pathol. 36, 548–552

90Misselwitz, B., Schmitt-Willich, H., Ebert, W., Frenzel, T., and Weinmann, H. J. (2001) Pharmacokinetics of Gadomer-17, a new dendritic magnetic resonance contrast agent Magn. Reson. Mater. Phys., Biol. Med. 12, 128–134

91Nazemi, A., Martínez, F., Scholl, T. J., and Gillies, E. R. (2012) Biodegradable dendritic polymersomes as modular, high-relaxivity MRI contrast agents RSC Adv. 2, 7971–7973

92Klemm, P. J., Floyd, W. C., Smiles, D. E., Fréchet, J. M. J., and Raymond, K. N. (2012) Improving T<sub>1</sub> and T<sub>2</sub> magnetic resonance imaging contrast agents through the conjugation of an esteramide dendrimer to high-water-coordination Gd(III) hydroxypyridinone complexes Contrast Media Mol. Imaging 7, 95–99

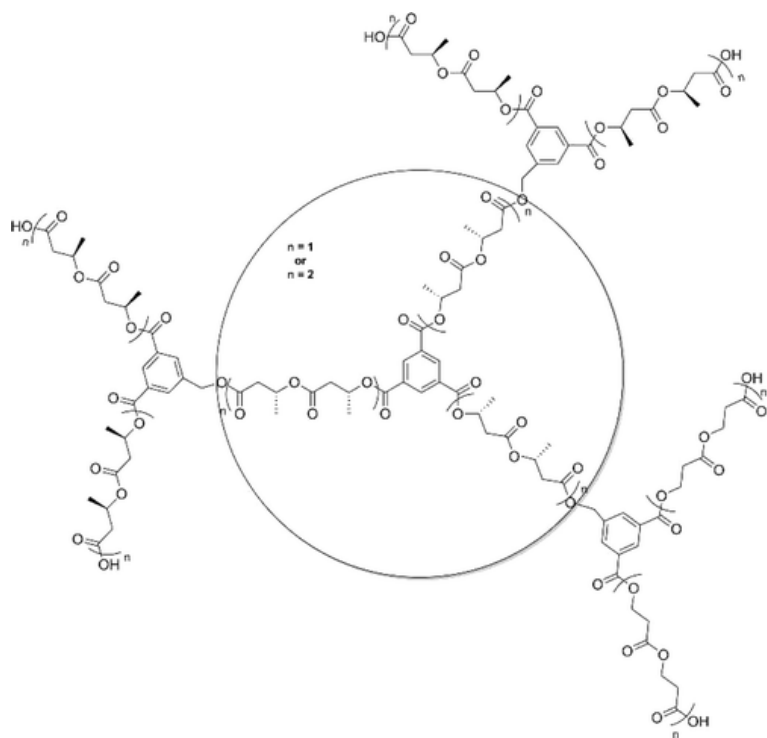
93Li, T., Qian, Y., Ye, M., Tang, J., Hu, H., and Shen, Y. (2014) Synthesis and properties of a biodegradable dendritic magnetic resonance imaging contrast agent Chin. J. Chem. 32, 91–96

94Klemm, P. J., Floyd, W. C., Andolina, C. M., Fréchet, J. M. J., and Raymond, K. N. (2012) Conjugation to biocompatible dendrimers increases lanthanide T<sub>2</sub> relaxivity of hydroxypyridinone complexes for magnetic resonance imaging *Eur. J. Inorg. Chem.* 2012, 2108– 2114

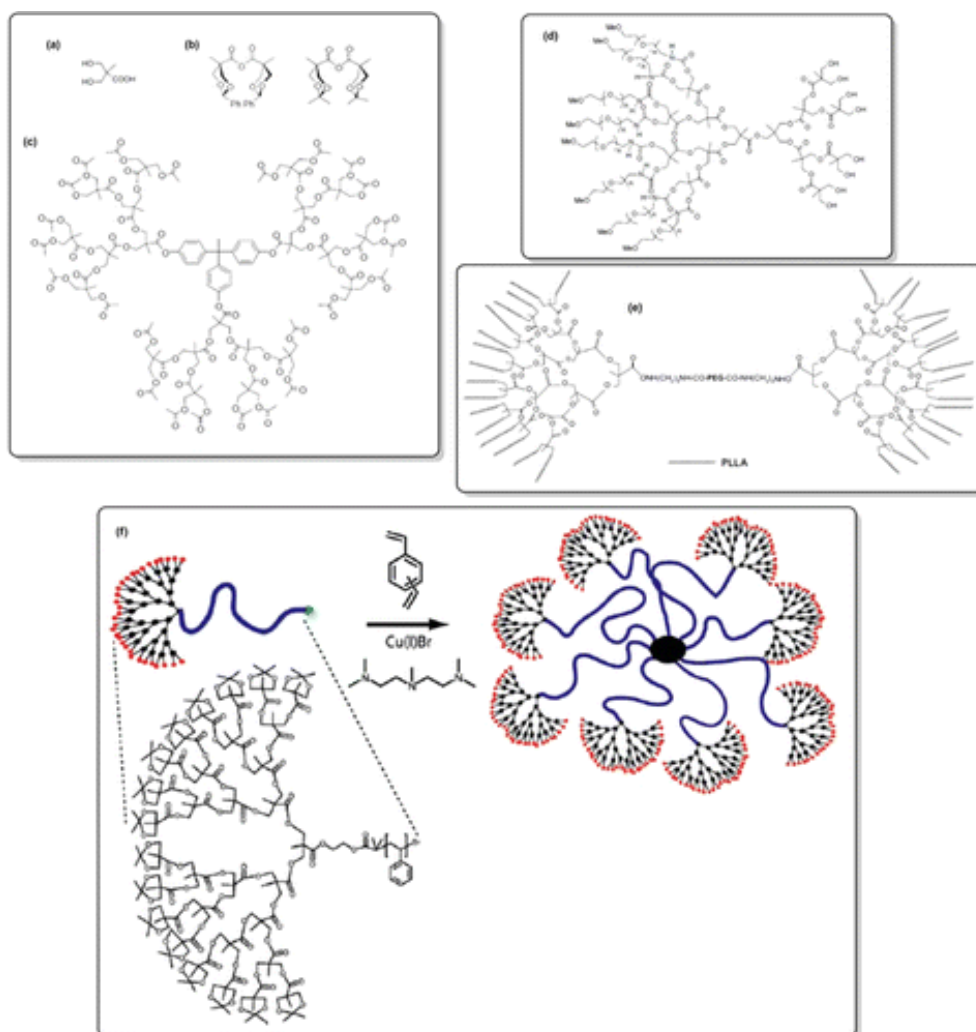
95Floyd, W. C., Klemm, P. J., Smiles, D. E., Kohlgruber, A. C., Pierre, V. C., Mynar, J. L., Fréchet, J. M. J., and Raymond, K. N. (2011) Conjugation effects of various linkers on Gd(III) MRI contrast agents with dendrimers: optimizing the hydroxypyridinonate (HOPO) ligands with nontoxic, degradable esteramide (EA) dendrimers for high relaxivity *J. Am. Chem. Soc.* 133, 2390– 2393

96Sosnovik, D. E. and Weissleder, R. (2007) Emerging concepts in molecular MRI *Curr. Opin. Biotechnol.* 18, 410

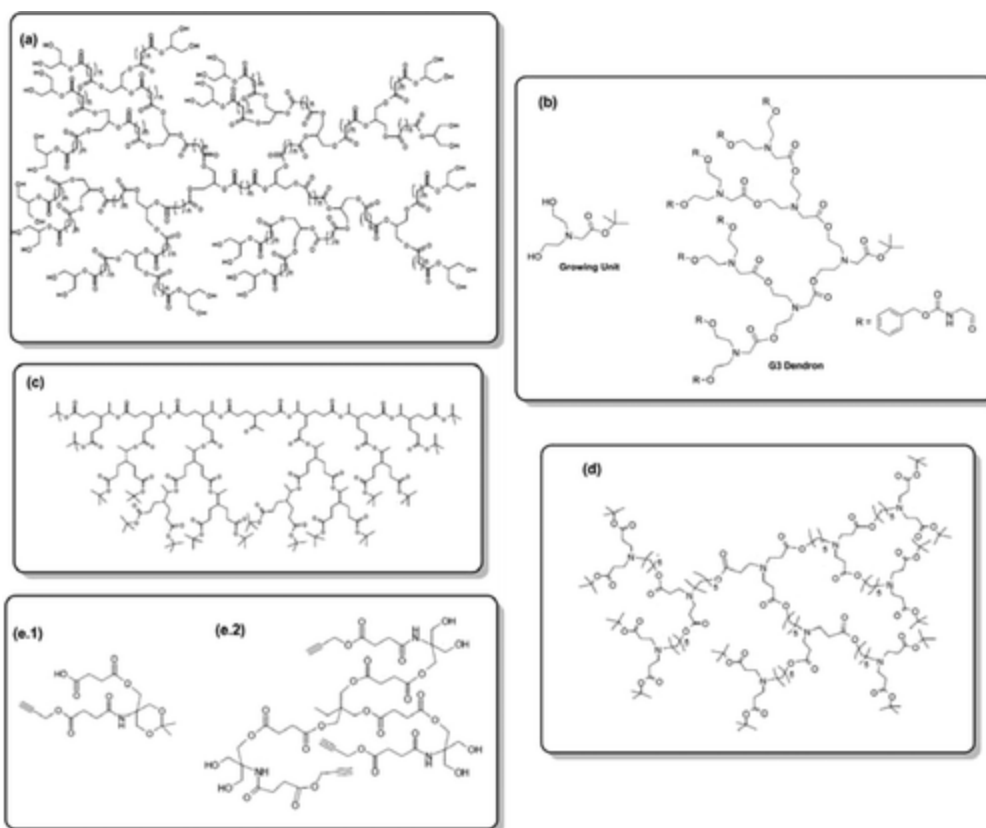
97Gallez, B. and Swartz, H. M. (2004) In vivo EPR: when, how and why? *NMR Biomed.* 17, 223– 225



**Figure 1.** Generation 1 (G1) of the first published biodegradable dendrimer (the dendrimer core is depicted within the circle).

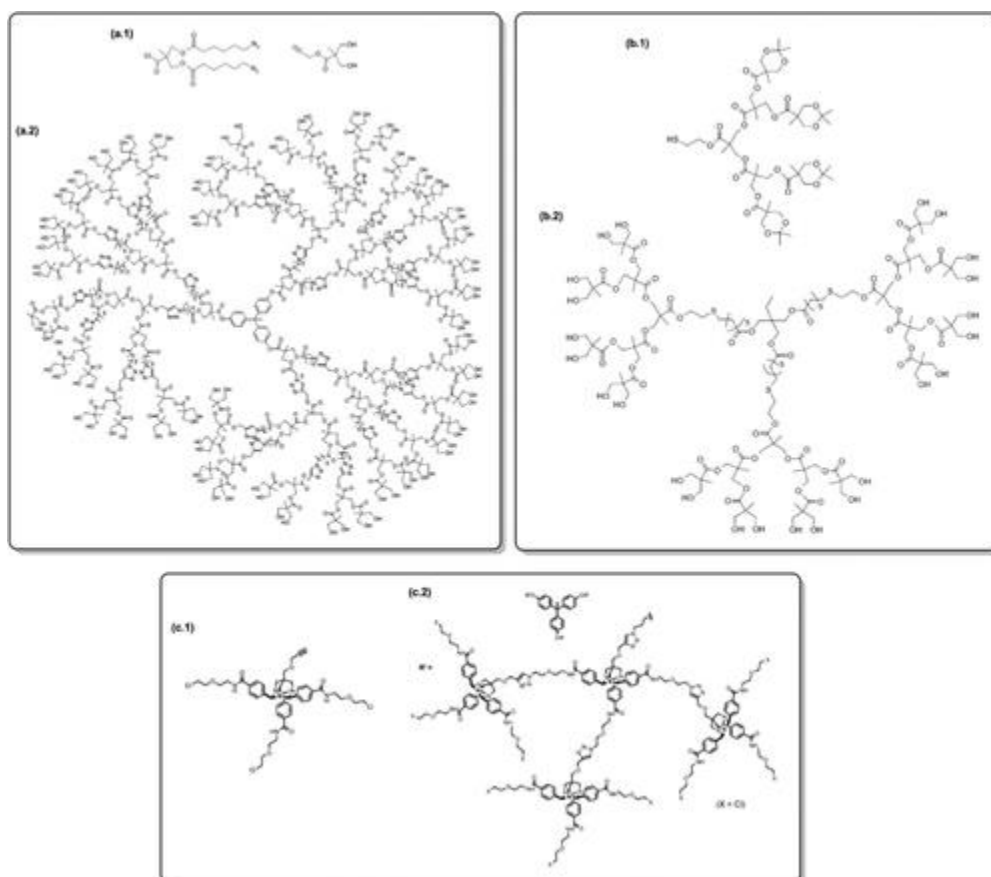


**Figure 2.** Bis-HMPA based dendrimers (a) 2,2-bis(hydroxymethyl)propanoic acid (bis-HMPA). (b) Protected anhydrides for the synthesis of bis-HMPA based dendrimers. (c) Generation 3 (G<sub>3</sub>) of the first synthesized bis-HMPA dendrimer. (d) "Bow-tie" dendrimer. (e) Structure of dumbbell-shaped triblock copolymer (adapted from Biodegradable comb-dendritic triblock copolymers consisting of poly(ethylene glycol) and poly(L-lactide): Synthesis, characterizations, and regulation of surface morphology and cell responses, Gong, F., Cheng, X., Wang, S., Wang, Y., Gao, Y., Cheng, S. *Polymer* 2009, 50, 2775–2785 reprinted with permission from Elsevier). (f) Synthesis of Generation 5 functionalized Core Cross-linked Star (CCS) polymers via the "arm first" approach. (Reproduced from *Macromolecules* 2007, 40, 7855–7863 with permission from The Royal Society of Chemistry).

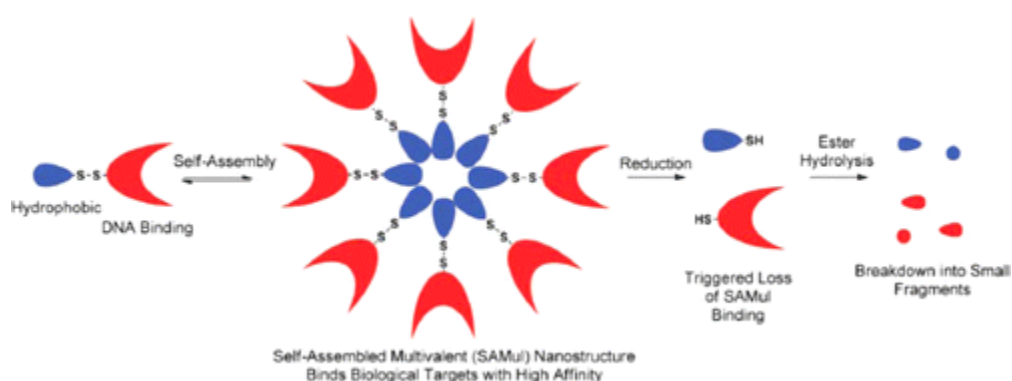


**Figure 3.** (a) Generation 3 (G3) of a polyester "biodendrimer". (b) Growing unit and Generation 3 (G3) of biodegradable ester-amine dendrons. (c) Generation 3 (G3) of the acetoacetate tert-butyl acrylate derived dendrimer. (d) Second generation (G2) of amine-containing polyester dendrimer. (e) Bifunctional dendrimers: (e.1) AB<sub>2</sub>C dendron, (e.2) first generation (G<sub>1</sub>) of the bifunctional dendrimer with three alkynes and six hydroxyls.

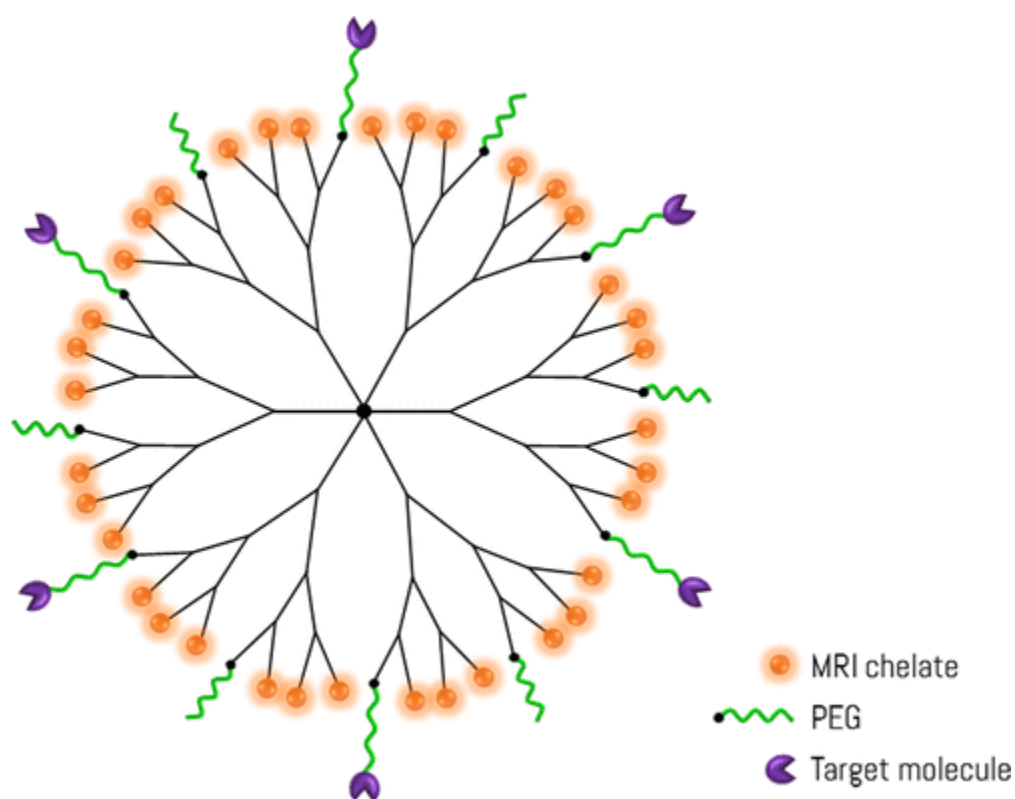




**Figure 4.** (a) Alternating polyester dendrimer: (a.1) AB<sub>2</sub> dendrons, (a.2) fourth generation (G<sub>4</sub>) dendrimer resulting from accelerated synthesis. (b) Alternating polyester dendrimer: (b.1) macrothiol-dendrion bearing latent hydroxyls, (b.2) hydroxyl-terminated dendrimer based on macrothiol-dendrons. (c) Degradable TAA-based dendrimers: (c.1) degradable TAA monomer, (c.2) generation 2 (G<sub>2</sub>) of TAA-based dendrimers.



**Figure 5.** Schematic representation of the concept of triggered loss of multivalent binding (reproduced from Org. Biomol. Chem.2014, 12, 446–455 with permission of The Royal Society of Chemistry).



**Figure 6.** Schematic representation of a PEGylated and targeted dendritic contrast agent.

**Table 1.** Biodegradable Dendrimer-Based MRI Contrast Agents

dendrimer	generation	chelate	ionic relaxivity <sup>a</sup> (mM <sup>-1</sup> s <sup>-1</sup> )	author/ref
PE Dendron coupled to PCL-PEO polymersome	3	Gd(III)-DTPA	$T_1 = 26.1^*$	Nazemi et al. <a href="#">(91)</a>
PEG-conjugated EA dendrimer	-	Gd(III)-TACN-bis(1-Me)-3,2-HOPO-TAM-ethylamine	$T_1 = 31$ ; $T_2 = 52^{**}$	Klemm et al. <a href="#">(92)</a>
PE dendrimer	0–3	Gd(III)-DTPA	$T_1 = 11.7^{***}$	Ye et al. <a href="#">(14)</a>
FA PEG-conjugated PE dendrimer	2	Gd(III)-DTPA	$T_1 = 17.1^{***}$	Ye et al. <a href="#">(89a)</a>
PEG-conjugated PE dendrimer	1	Gd(III)-DTPA	$T_1 = 10.2^{***}$	Li et al. <a href="#">(93)</a>
PEG-conjugated PE dendrimer	2	Gd(III)-DTPA	$T_1 = 10.7^{***}$	Li et al. <a href="#">(93)</a>
PEG-conjugated PE dendrimer	4	Gd(III)-DTPA	$T_1 = 15.6^{***}$	Li et al. <a href="#">(93)</a>
PE dendrimer	5	Gd(III)-DTPA	$T_1 = 17.5^{***}$	Li et al. <a href="#">(93)</a>

<sup>a</sup> Measured at \*20 MHz and 25 °C, \*\*60 MHz and 37 °C, \*\*\*0.52 T and 32 °C.



**INSTITUTO  
DE INVESTIGAÇÃO  
E INOVAÇÃO  
EM SAÚDE**  
UNIVERSIDADE  
DO PORTO

Rua Alfredo Allen, 208  
4200-135 Porto  
Portugal  
+351 220 408 800  
[info@i3s.up.pt](mailto:info@i3s.up.pt)  
[www.i3s.up.pt](http://www.i3s.up.pt)